

Nanomagnetism

Part 2: Magnetic Nanoparticle Applications

Sara A. Majetich

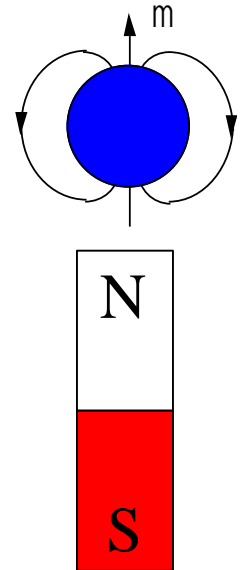
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Applications of Magnetic Nanoparticles

1. Intro and key nanoparticle properties
2. Magnetic manipulation
3. Magnetic sensing
4. Magnetic hyperthermia
5. Magnetic Particle Imaging (MPI)
6. Magnetic Resonance Imaging (MRI)



Where Do You Have Magnetic Nanoparticles?

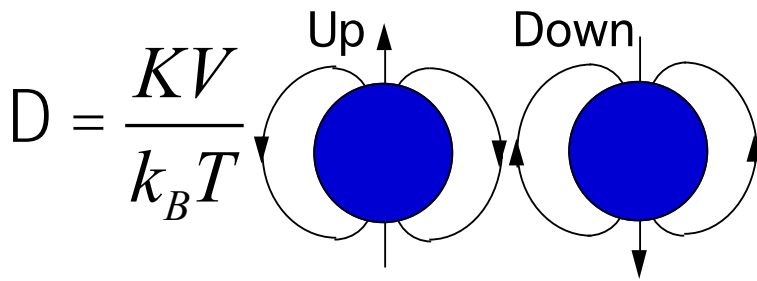


Magnetic Hard Disk Media



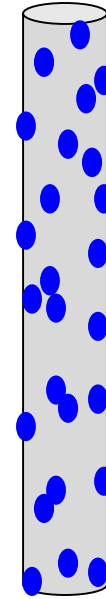
Magnetic stripe
(data on tracks 1 & 2)

Credit Card Magnetic Stripe

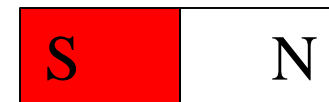


Monodomain, not superparamagnetic

Where Do You Have Magnetic Nanoparticles?



Iron oxide nanoparticles grown in cellulose fibers used in \$ as anti-counterfeiting measure



Use a NdFeB magnet to exert a force

Have You Ever *Eaten* Magnetic Particles?

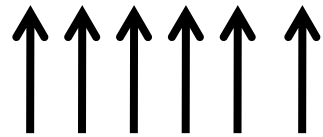
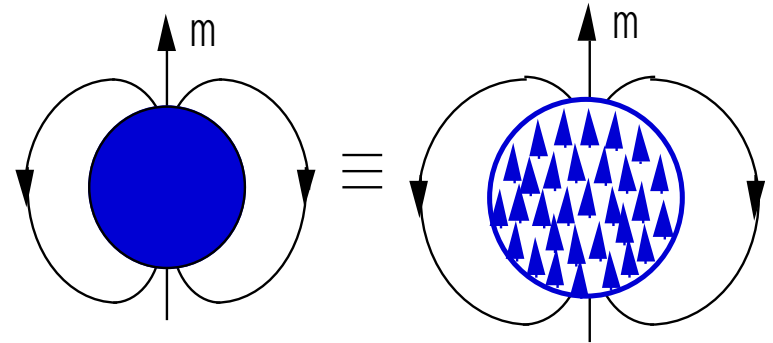
Iron-fortified cereals have $< 45 \mu\text{m}$ food-grade Fe particles

- dissolve in stomach acids so the Fe can be absorbed
- $\text{Fe}(\text{PO})_4$ and FeSO_4 also used for fortification



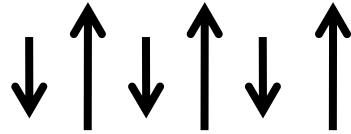
Magnetic Particle Review

Monodomain: no domain walls, so acts like giant spin



Ferromagnet

Fe



Ferrimagnet

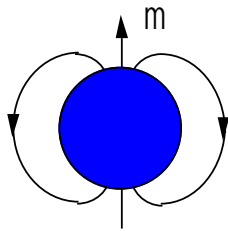
γ -Fe₂O₃, Fe₃O₄

Superparamagnet: monodomain particle where there are rapid fluctuations in the magnetization *direction*

Important **Magnetic** Features

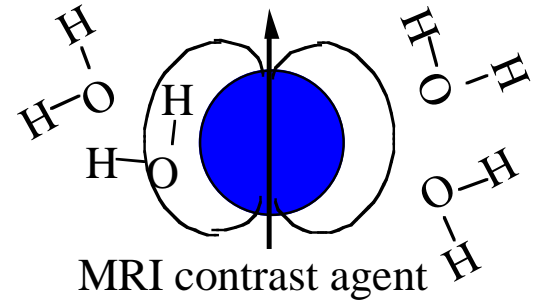
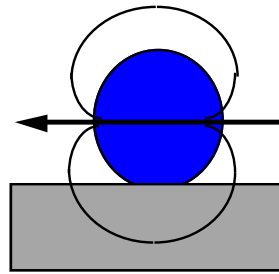
Magnetic moment

$$\mu = M_s V$$



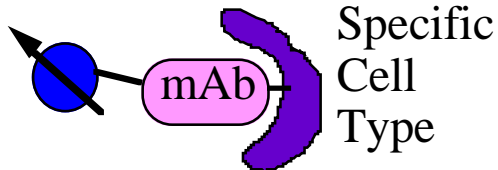
Generate a Magnetic Field $H \sim \mu$

Magneto-resistive Sensor



Move in response to a magnet

$$F = \mu \cdot \nabla H$$



$$D = \frac{KV}{k_B T}$$

Thermal stability enables information storage

Important **Magnetic** Features for Specific Applications

Application	Generates Field	Moves in Field	Magnetically Metastable
Magnetic Manipulation	N	Y	N
Magnetic Sensing	Y (for detection)	N	Y(over measurement time)
Magnetic Hyperthermia	N	N	N (for DC), but Y (for AC field used)
Magnetic Particle Imaging	Y (for detection)	N	N
Magnetic Resonance Imaging	Y	N	N
Ferrofluids	Y (to form chains)	Y	N
Magnetic Recording Media	Y (for reading)	N	Y (for a long time)

Biomedical Applications of Magnetism

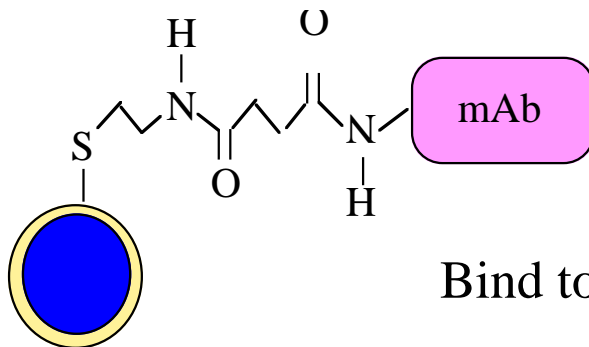
- Magnetic fields disturb normal biological function much less than electric fields.
- Use the power of nanomagnetism as a tool, with some constraints, in complex biological systems



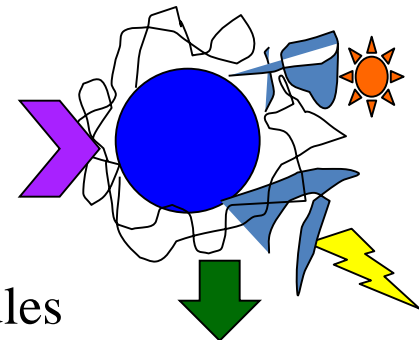
Requirements *Beyond* Magnetism

Toxic? Mutagenic? Carcinogenic?

- Stricter requirements for **in vivo** than for **in vitro**
- **LD50** (Median lethal dose, per kg of body mass) – *In vitro* cell exposure test – determine exposure for half of cells to die
- Co, Ni demonstrated carcinogens
- **Biocompatible** (no immunoresponse) **vs. biodegradable** (body excretes)
- **Only superparamagnetic iron oxide** (Endorem^R, Feridex^R, Resovist^R) has in vivo approval



Bind to targeting molecules



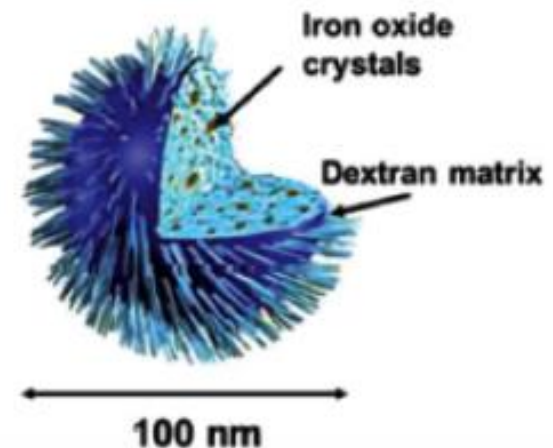
Importance of Particle Coatings

Coatings are used to **enhance dispersion stability**, to **functionalize for selective binding**, and (for in vivo use) to **prevent rapid removal**

- The Reticulo-endothelial system (RES) recognizes “intruders”, coats with proteins, so they are recognized and removed by macrophages (opsonization)

- Coating magnetic nanoparticles with “friendly” molecules essential

(Cindi Dennis, NIST)

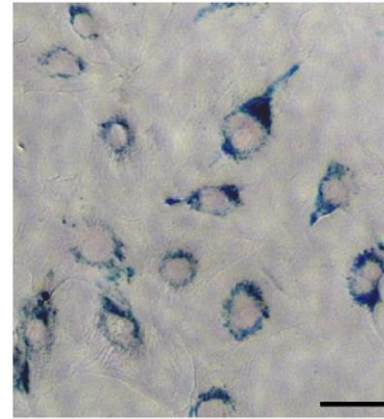
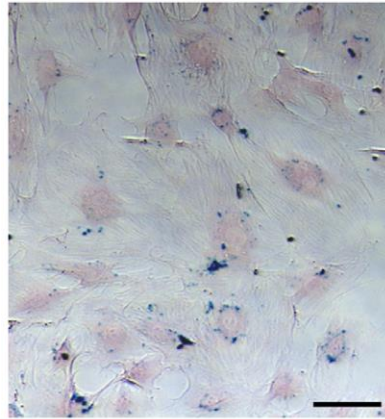


C. C. Berry and A. S. G. Curtis, J. Phys. D: Appl. Phys. **36** R198-R206 (2003).

V. P. Torchilin, Amer. Assoc. Pharmaceut. Sci. Journal **9** E128-E147 (2007).

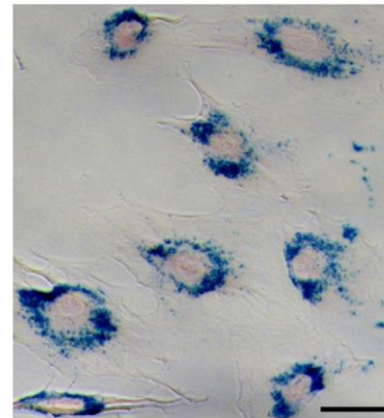
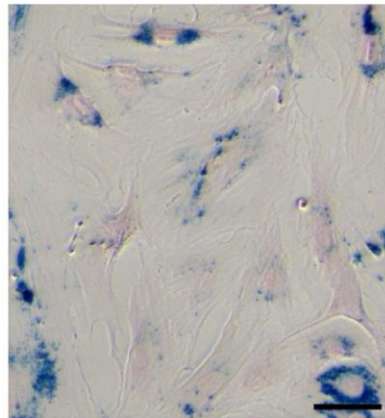
Coating and Stem Cell Uptake Efficiency

Endorem^R
(dextran-coated)



Poly(lysine)-
coated
Endorem^R

Uncoated $\gamma\text{-Fe}_2\text{O}_3$

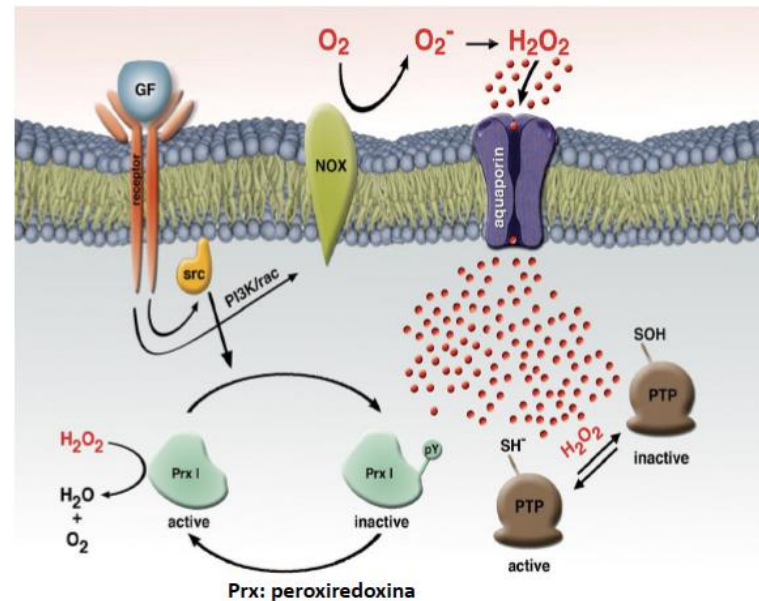
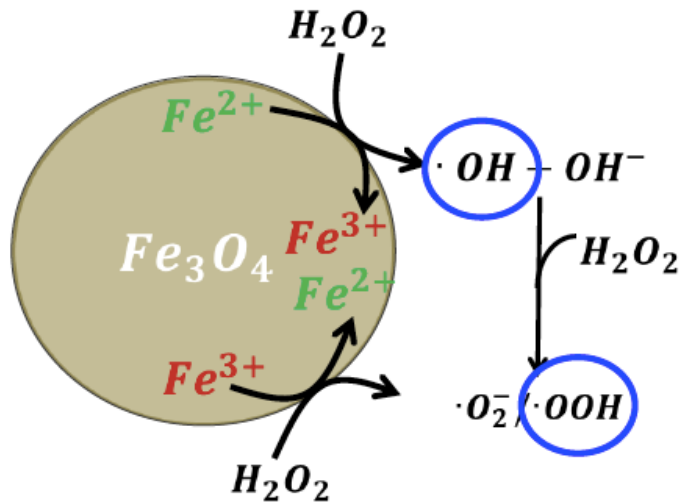


D-mannose-
coated $\gamma\text{-Fe}_2\text{O}_3$

Prussian Blue stain for Fe, Optical microscopy

Fe²⁺, Fe³⁺ Catalysis at NP Surfaces

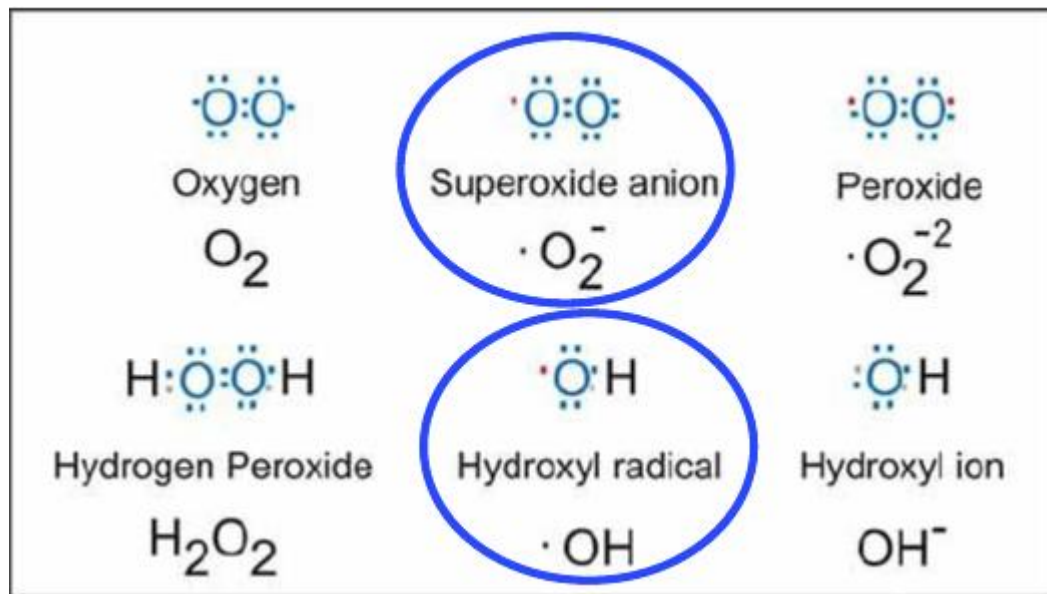
Cells regulate the balance between creation and destruction of free radicals, but NPs can shift the balance



Roberto Zysler, INN Bariloche

Reactive Oxygen Species (ROS)

60% of DNA damage due to X-rays from ROS (75% from $\bullet\text{OH}$)



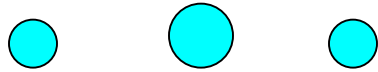
$\bullet\text{OH}$ can cause oxidative stress and damage

If Fe^{2+} can alter the ROS concentration, are Mn ferrite ($\text{Mn}^{2+}\text{Fe}^{3+}\text{O}_4$) NPs better than Magnetite, ($\text{Fe}^{2+}\text{Fe}^{3+}\text{O}_4$)? Is $\gamma\text{-Fe}_2\text{O}_3$ (no Fe^{2+} , but lower M_s) better?

Particle Dispersion

- If particles form large agglomerates in biological media, they could cause heart attacks or strokes in living organisms
- What happens in biological media? (Phosphate Buffered Saline solution used is equivalent to 154 mM NaCl --- need steric stabilization)
- Primary particle size from TEM, clustering from dynamic light scattering (DLS)

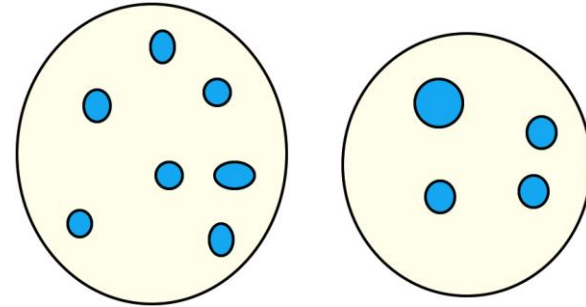
Two Main Types of Nanoparticles



Fe_3O_4 or $\gamma\text{-Fe}_2\text{O}_3$; ~ 10 nm

Individual particles

MRI, hyperthermia,
ferrofluids



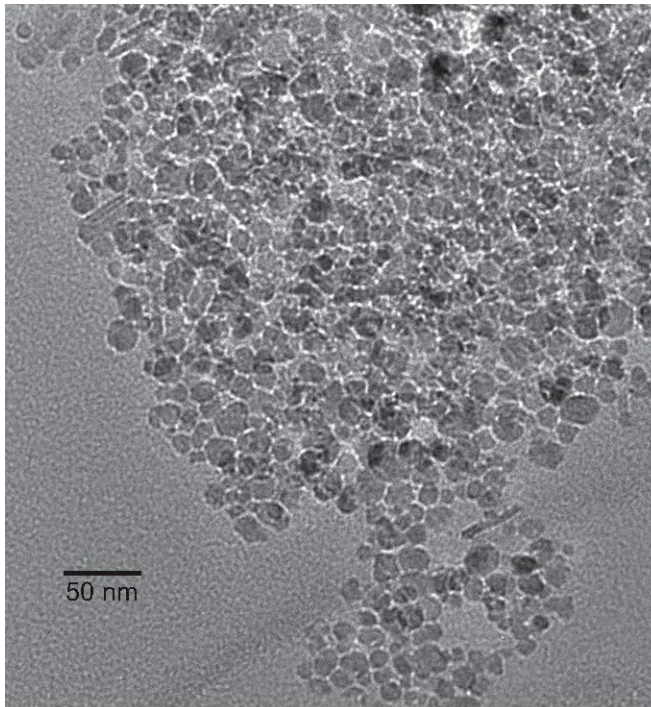
Magnetic beads with many
particles bound in a polymer
matrix; $\sim 0.5\text{-}10$ μm

Magnetic separation, sensing

Coatings important for both types

Superparamagnetic

Coprecipitation or Hydrothermal Synthesis

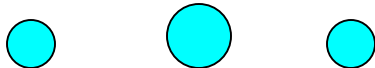


Mix Fe (II), Fe (III) salts in aqueous solution, add base

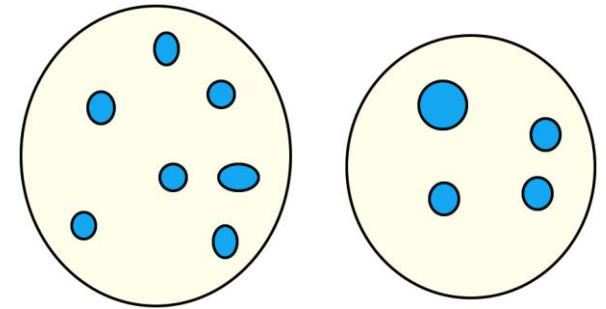
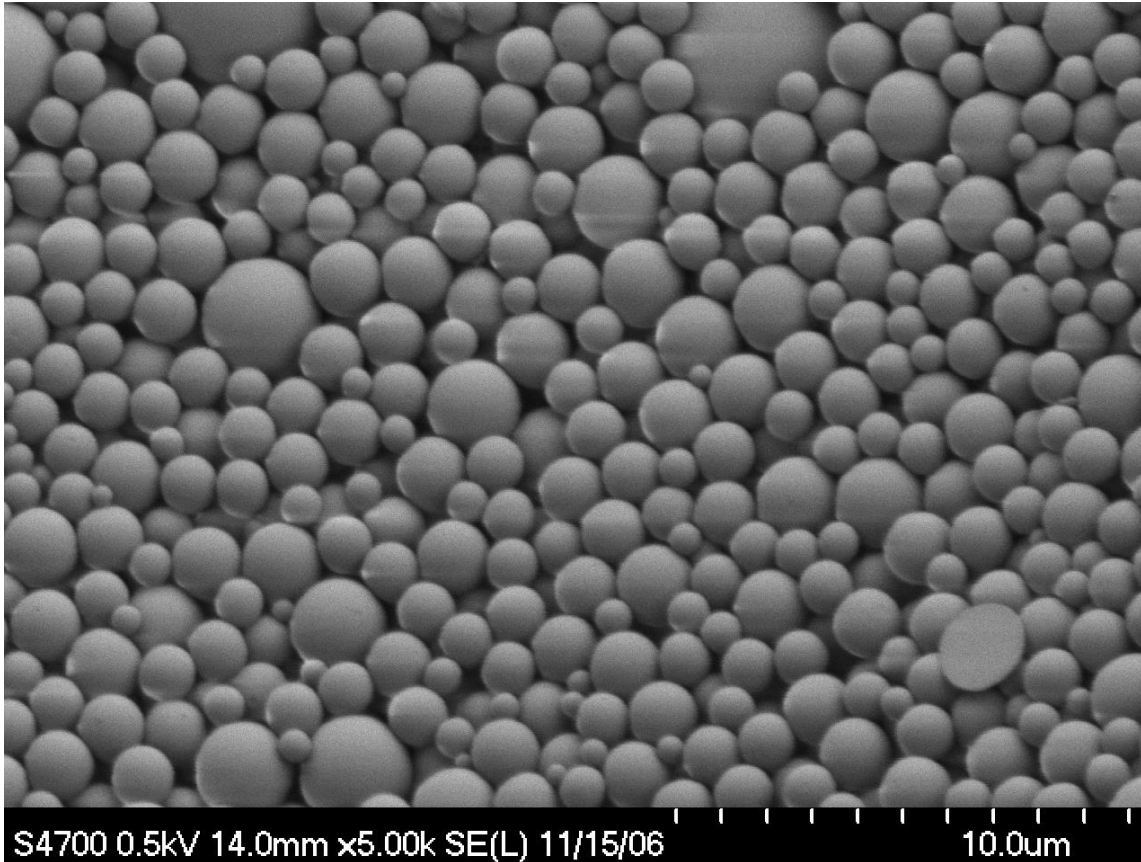
Easy, all reagents can be biocompatible, makes ~10 nm particles, 20-50 emu/g

GMP: Good manufacturing practices

R. Massart and V. Cabuil, J. Chem. Phys. **84** 967 (1987).



Magnetic Beads

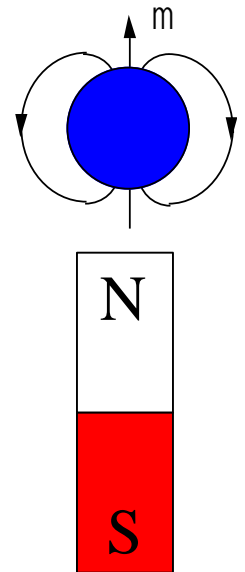


Commercially
available (Dyna,
Miltenyi,...)

H. Zhao, K. Saatchi, U. O. Hafeli, J. Magn. Magn. Mater. **320**, 1356 (2009).

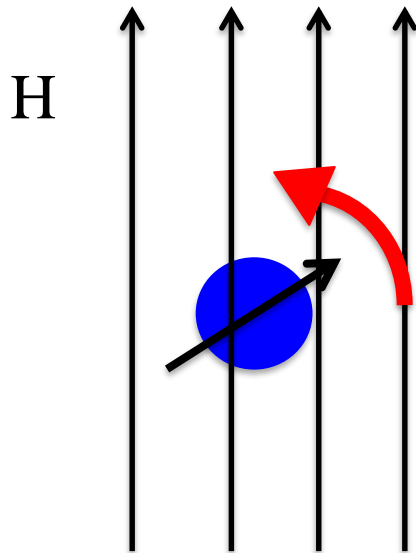
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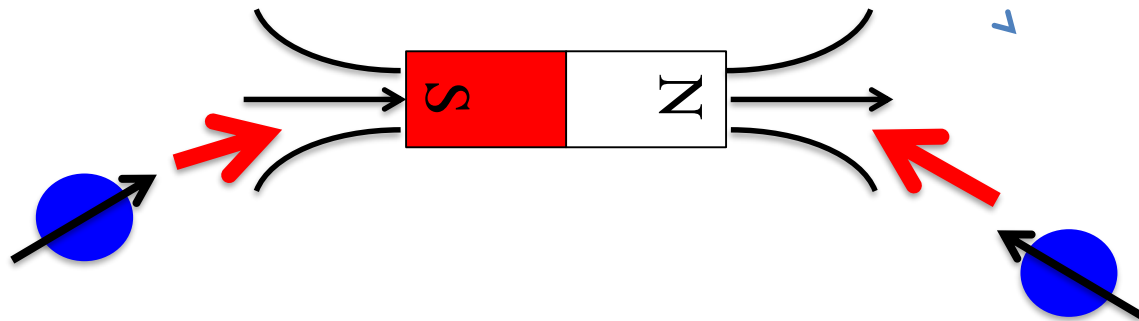
2 Kinds of Magnetically Controlled Motion

Particle moment μ
rotates in a uniform field



Torque $\tau = \mu \times H$

Spatially varying translates
particles toward poles



Force $F = \mu \cdot \nabla H$

Torque on Vortex Discs for Cancer Treatment

nature
materials

ARTICLES

PUBLISHED ONLINE: 29 NOVEMBER 2009 | DOI: 10.1038/NMAT2591

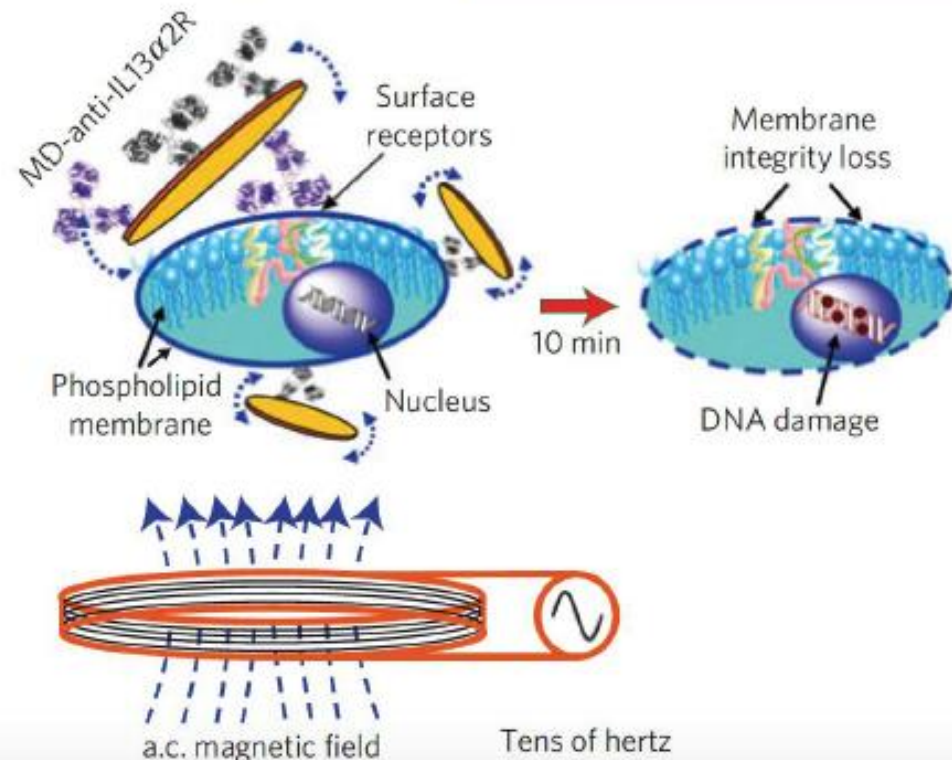
Biofunctionalized magnetic-vortex microdiscs for targeted cancer-cell destruction

Dong-Hyun Kim¹, Elena A. Rozhkova^{2*}, Ilya V. Ulasov³, Samuel D. Bader^{1,2}, Tijana Rajh², Maciej S. Lesniak³ and Valentyn Novosad^{1*}

Torque $\tau = \mathbf{m} \times \mathbf{H}$

Oscillation of the disks induces programmed cell death

Kim, Nature Mater. 9 165, (2010)



Magnetophoresis

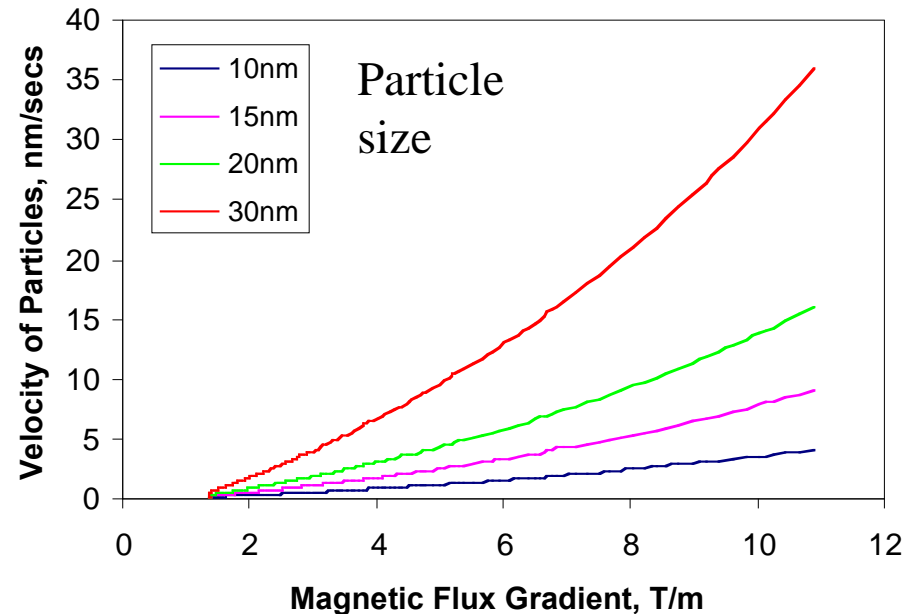
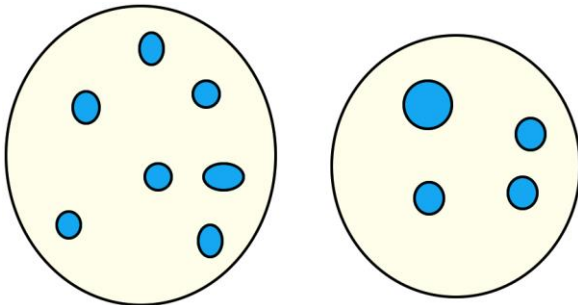
- Drag forces large for small particles, relative to magnetic force

$$F_{drag} = 6\rho h r_{hyd} \quad \vec{F}_{mag} = (\vec{\mu} \cdot \nabla) \vec{B}$$

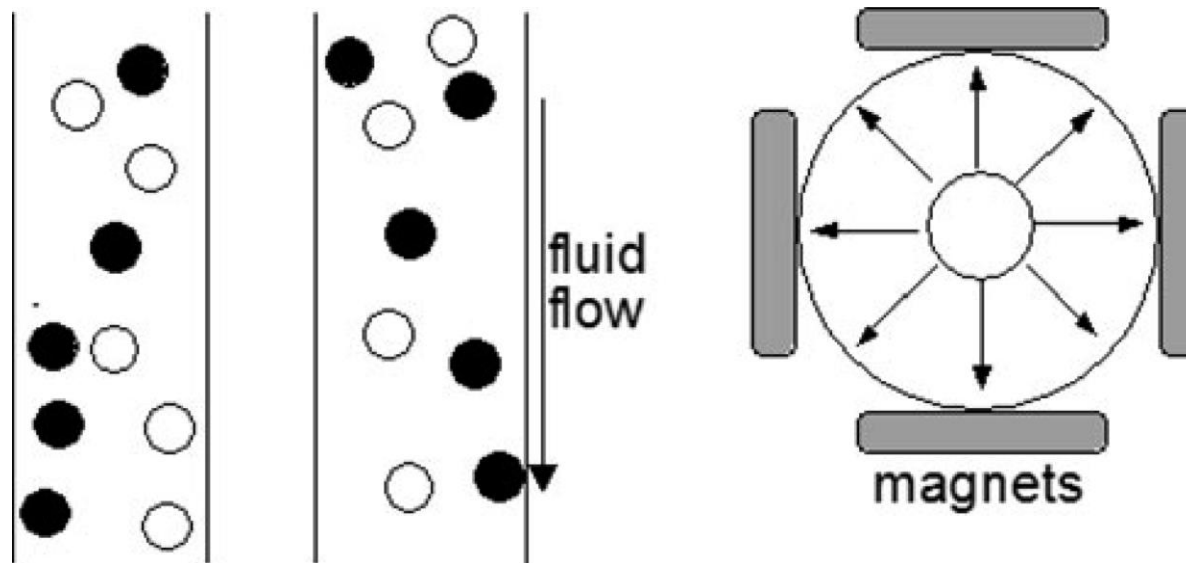
- Terminal velocity

$$u_{mag} = \frac{F_{mag}}{6\rho h r_{hyd}}$$

- Use magnetic beads



Quadrupole Magnetic Separation of Circulating Tumor Cells



F. Carpino, et al. J. Magn. Magn. Mater. 311 383 (2007).

S. Nagrath, et al., Nature 450 20 (2007) - < 10 cells/10 mL blood, Stage I breast cancer trial

Circulating Tumor Cells

Primary cancers shed cells even when tumor seems to be organ-confined, cause of metastasis

- E. Racila, et al., PNAS **95** 4589-4594 (1998).

Detection of tumor cells in blood could be used for diagnosis, choice of treatment, prognosis

Challenge: find 10 cancer cells among 5×10^9 red blood cells 5×10^6 white blood cells/mL of blood

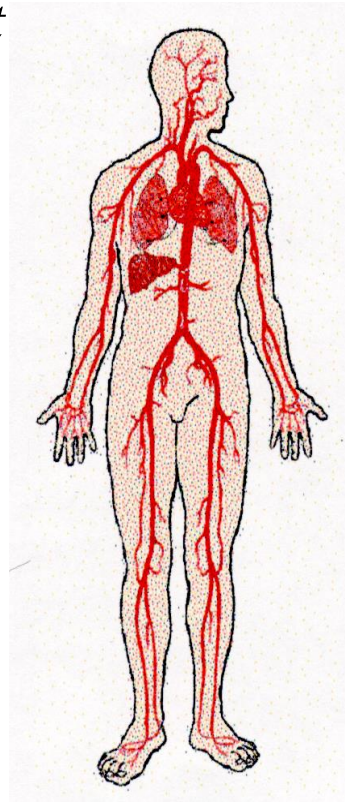
Targeting Magnetic Particles *in vivo*

No **magnetic** drug delivery *yet*

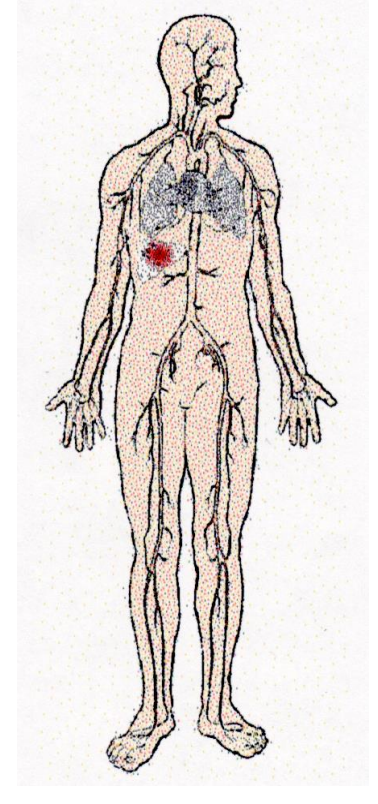
- Hard to guide to deep locations
- Can hold in place with external magnets

-Selective chemical binding very inefficient

-Best delivery by injection upstream of tumor



Systemic Therapy

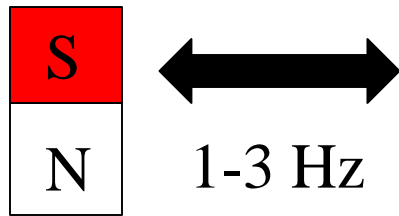
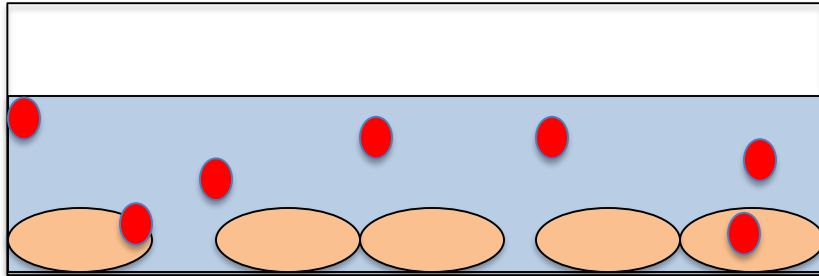


Drug Targeting

Urs Hafeli, Univ. of British Columbia

www.magneticmicrosphere.com/hafeli_lab

In Vitro Magnetic Gene Delivery



Oscillate field, stimulate endocytosis of 1 μm beads

- C. Plank, *Molec. Therapy* **6**, 106-112 (2002); *Nature Biotech.* **18**, 893-895 (2000).

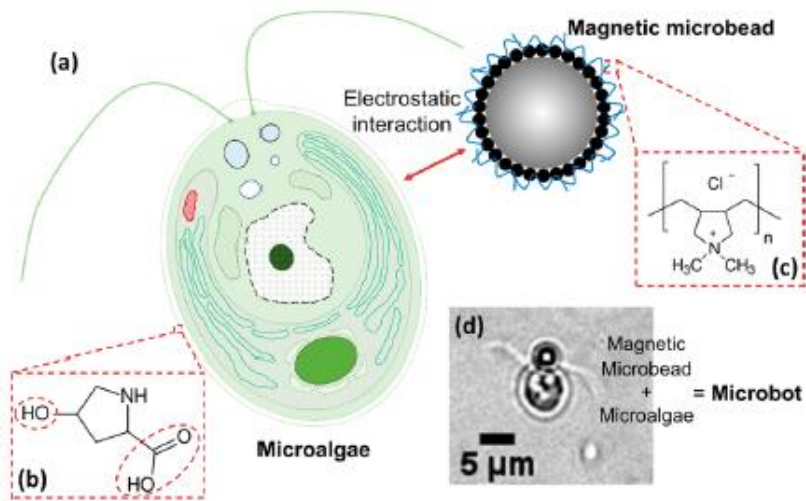
100 nm magnetic pts. coated with DNA coding for green fluorescence protein transfected

- S. C. McBain, et al., *Gene Therapy* **15**, 902 (2008)

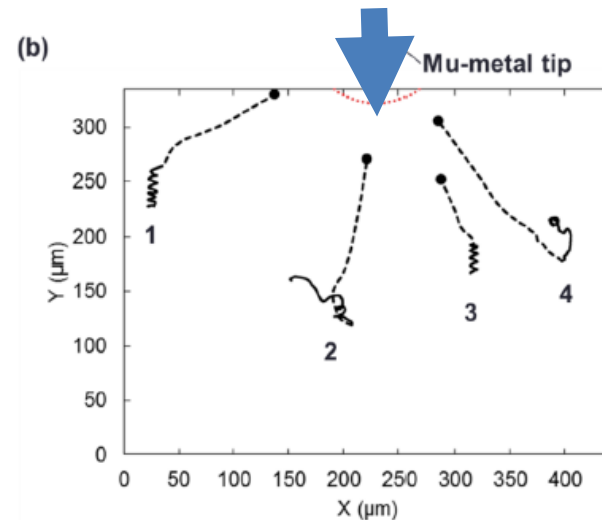
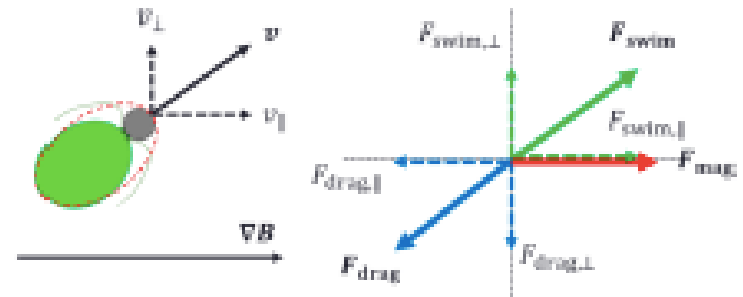
Feline fibrosarcoma trial – *J. Vet. Med. A Physiol. Pathol. Clinical Med.* **54** 599-606 (2007); human trials started in 2009

Magnetic Swimmers

Algae swim in response to field gradient



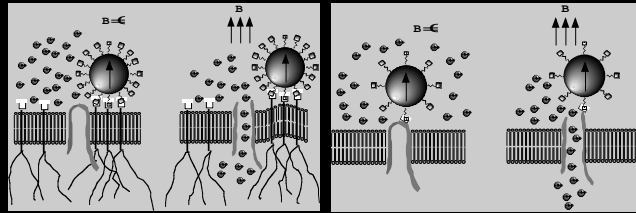
W. M. Ng, et al., *Langmuir* **34**, 7971 (2018)



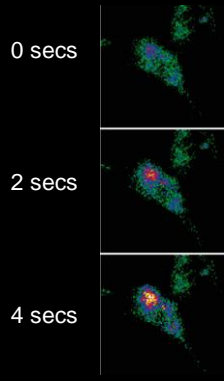
Helical trajectory due to flagella, but magnetic guidance

In Vitro Magnetic Control of Ion Channels

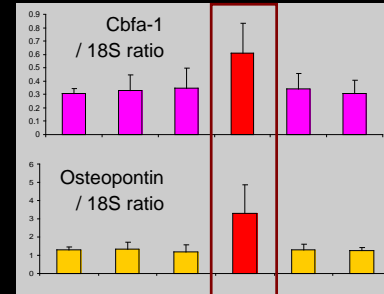
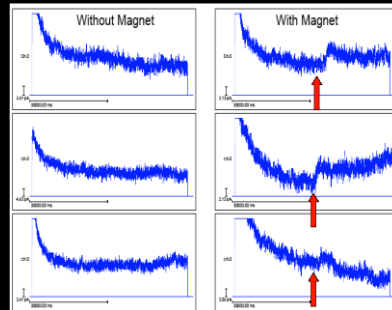
Magnetic ion channel activation (MICA) for Tissue Engineering & Regenerative Medicine



Basic Concept



Calcium (L) & Potassium (R) Ion Channel Activation



MSCs	✓	✓	✓	✓	✓	✓
Magnet		✓		✓		✓
TREK-1			✓	✓		✓
Ca channel Ab				✓		✓



Controlling stem cell differentiation in vitro (top) and in vivo (bottom)

Dobson, J (2008) *Nature Nanotechnology* 3: 139-143.
 Dobson, J & TG St. Pierre (1996) *Biochem. Biophys. Res. Commun.* 227:718-723.
 Cartmell et al. (2002) *IEEE Trans. NanoBiosci.* 1: 92-97.

Cartmell et al. (2005) *J Phys. Conf. Ser.* 17: 77-80.
 Dobson et al. (2006) *IEEE Trans. NanoBiosci.* 5: 173-177.
 Hughes, S, SC McBain, J Dobson, AJ El Haj (2008) *J. Royal Soc. Interface.* 5: 855-863.

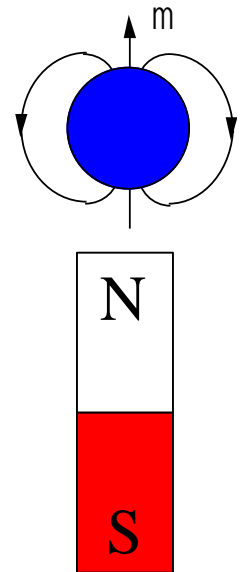


Jon Dobson, Keele University & Univeisty of Florida



Applications of Magnetic Nanoparticles

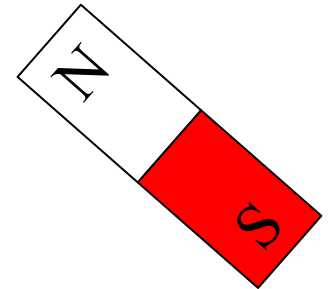
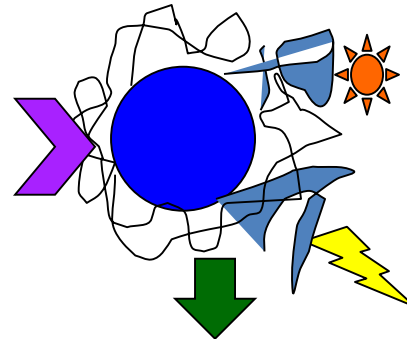
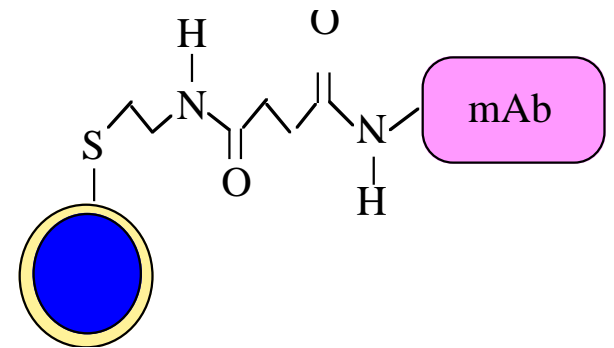
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Magnetic Sensing

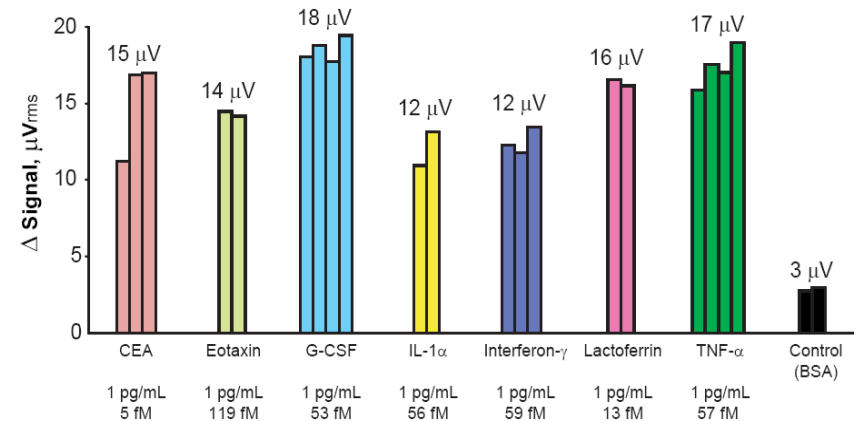
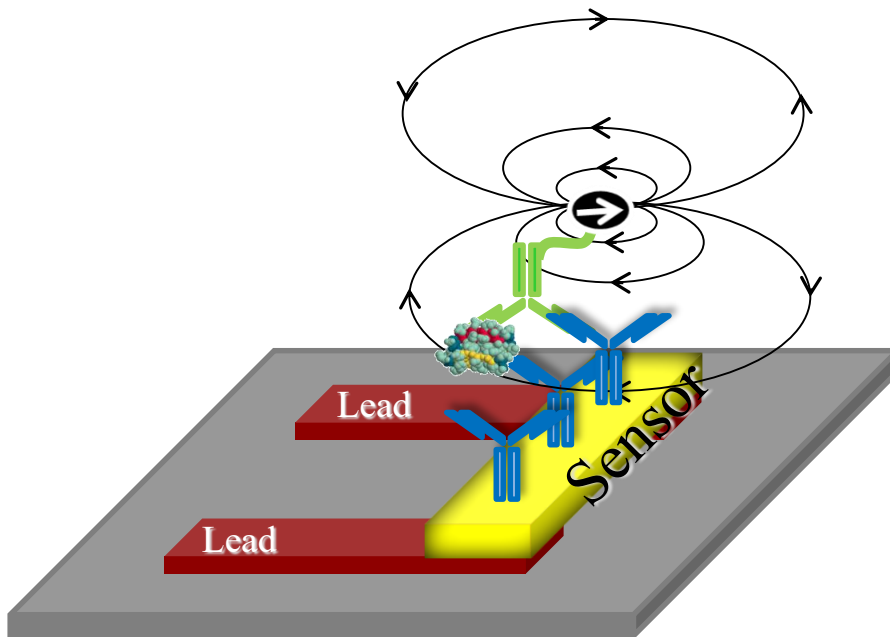
Use functionalized particles selectively bind analytes, then concentrate magnetically, then detect by:

- Magnetoresistance (GMR, TMR)
- Relaxometry (SQUID, fluxgate)



Magnetoresistive Sensing: Sandwich Assay

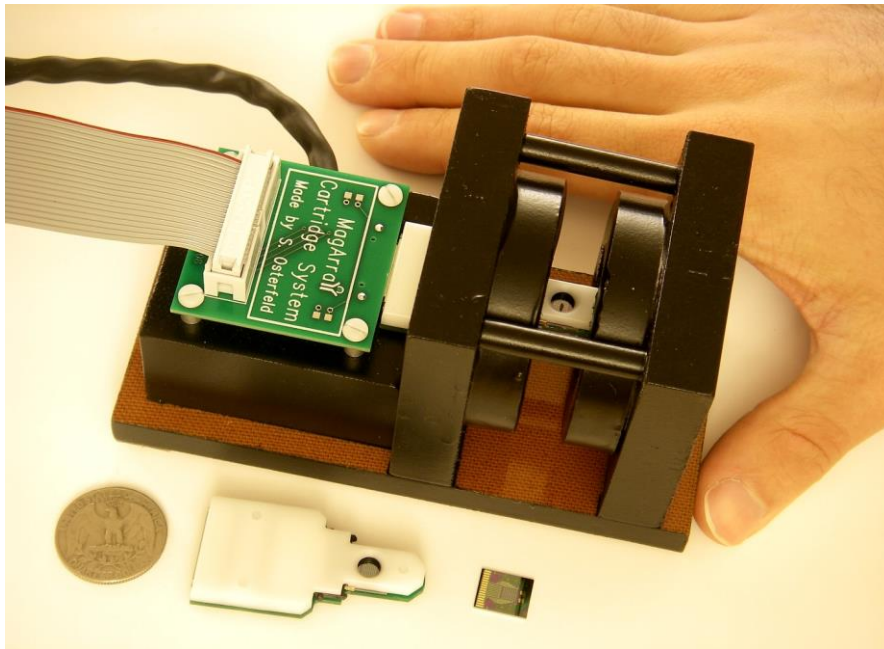
Need selective binding of analyte to NP + binding of NP to magnetic field sensor



S. J. Osterfeld, et al., PNAS, **105**, 20637, (2008)

Magnetic Blood Scanner for Cancer

- Uses magnetic beads to 'tag' proteins indicative of cancer and reading them out using magnetic sensors
- Higher sensitivity (1 picogram/mL) than conventional optical fluorescence assays, enabling earlier cancer detection.



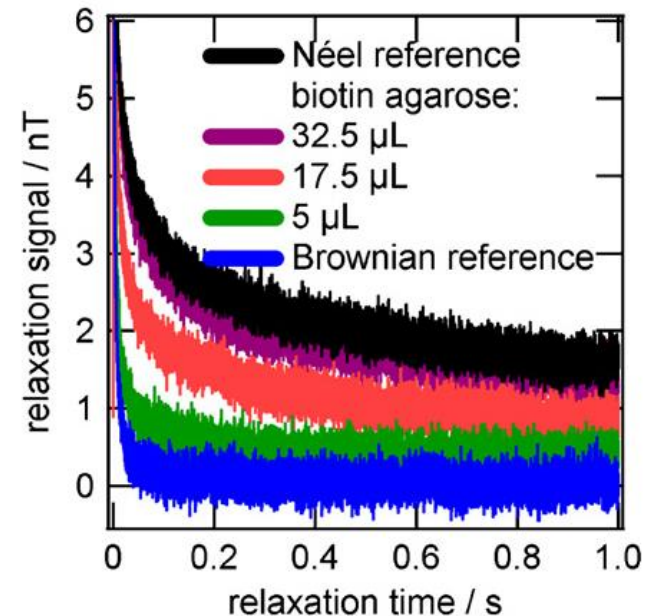
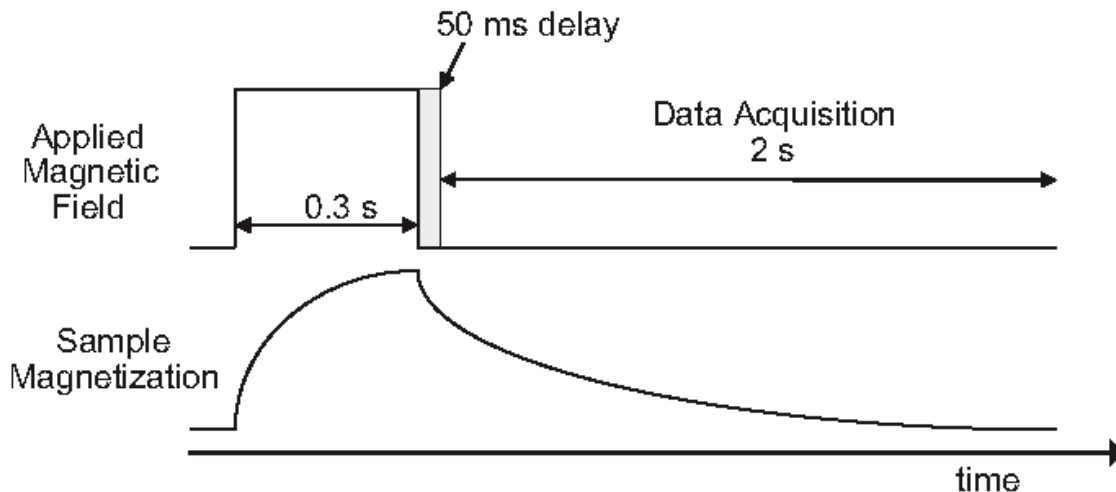
- Could be done quickly in a doctor's office

J. Choi, et al., Biosens. Bioelectron. 85, 1 (2016)

Relaxometry

3D sensor (microSQUID array) – **doesn't require particles to be within a few nm of a sensor surface**

Can do measurements on whole blood

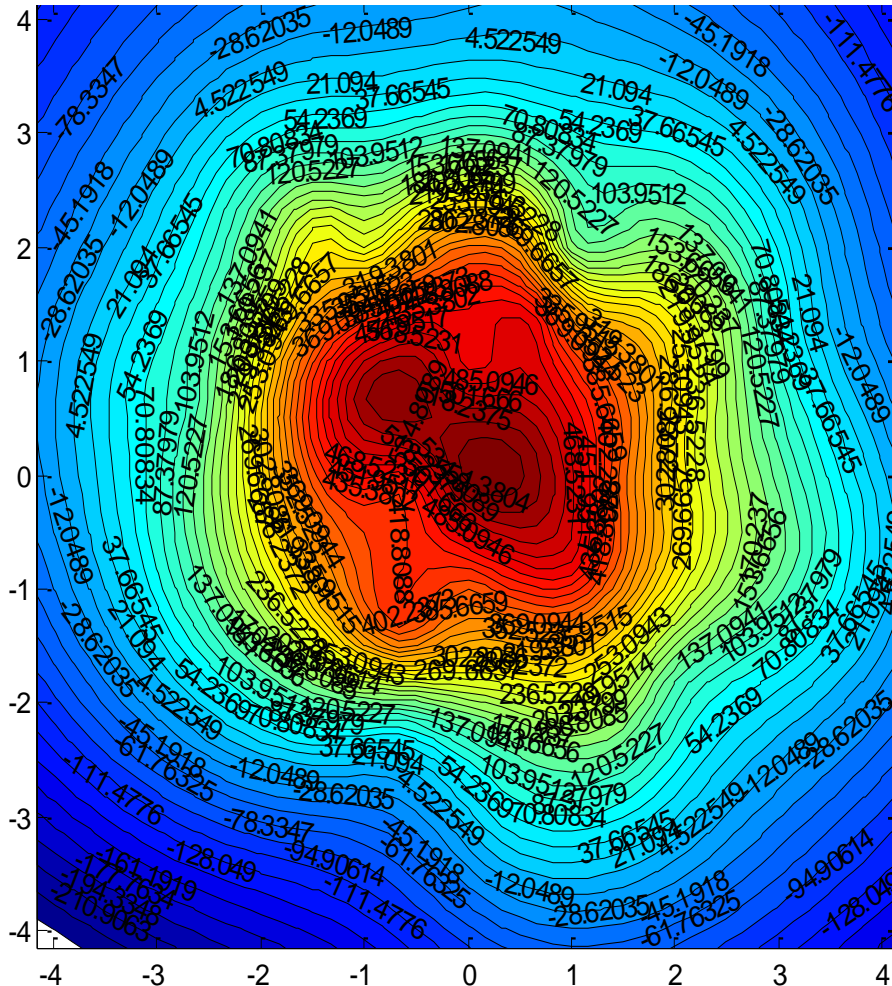


N. L. Adolphi, et al., *Contrast Media Molec. Imaging* **7**, 308 (2012).

D. Eberbeck, et al., *J. Magn. Magn. Mater.* **321** 1628-1631 (2009).

W. K. Peng, et al., *Nature Medicine* **20**, 1069 (2014).

SQUID Relaxometry Detection of Breast Cancer

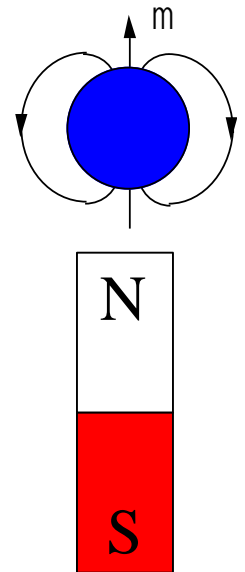


From mouse with two human breast tumors injected with anti-Her-2/neu labeled nanoparticles (Ocean Nanotech SHP-30)

Natalie L. Adolphi, Kimberly Butler, Debbie M. Lovato, Richard S. Larson (Univ. New Mexico) Trace E. Tessier, Howard C. Bryant, **Edward R. Flynn** (Senior Scientific, LLC)

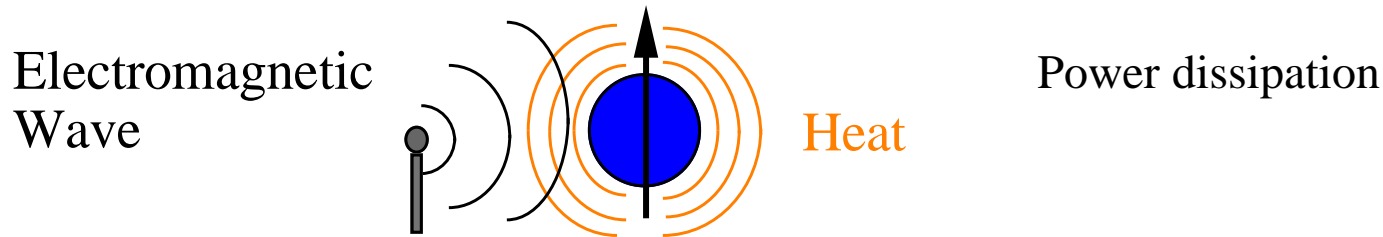
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Magnetic Hyperthermia

Dissipate Energy – Hyperthermia



Hyperthermia – $> 43^{\circ} \text{C}$ – cells more sensitive to chemotherapy drugs and radiation

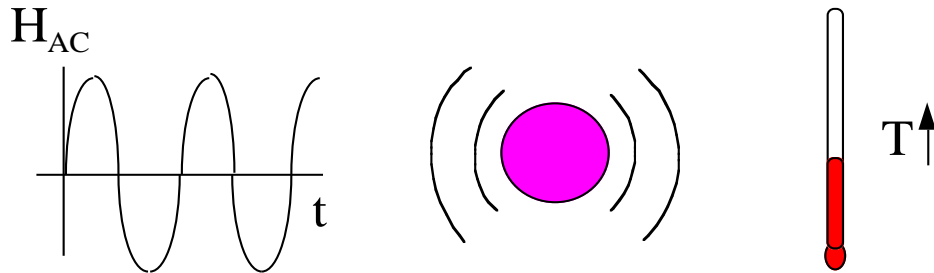
Ablation - $> 50^{\circ} \text{C}$ – kill cells directly by heating

Many types of hyperthermia: Direct, electrical, ultrasound, photothermal, magnetic (**where is magnetic hyperthermia superior?**)

Clinical trials: Charité Hospital Berlin, University College London

Magnetic Hyperthermia

Magnetic dissipation



$$\Delta U = -\mu_0 \oint M dH$$

$$P = f \Delta U = \mu_0 \pi \chi'' f H^2$$

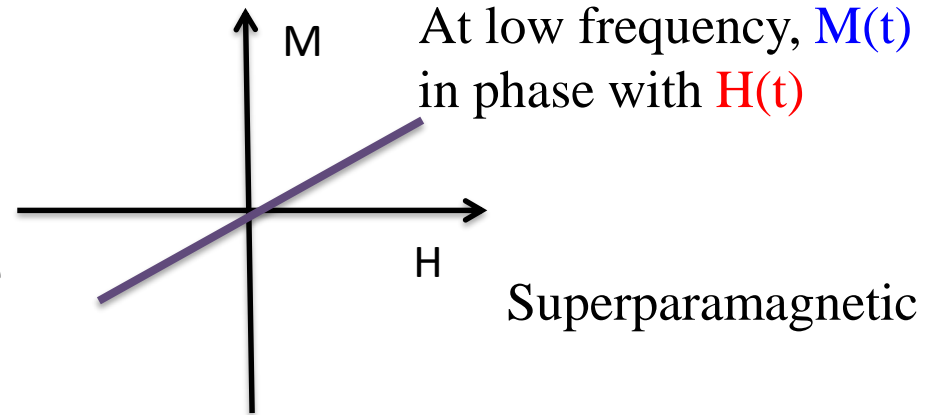
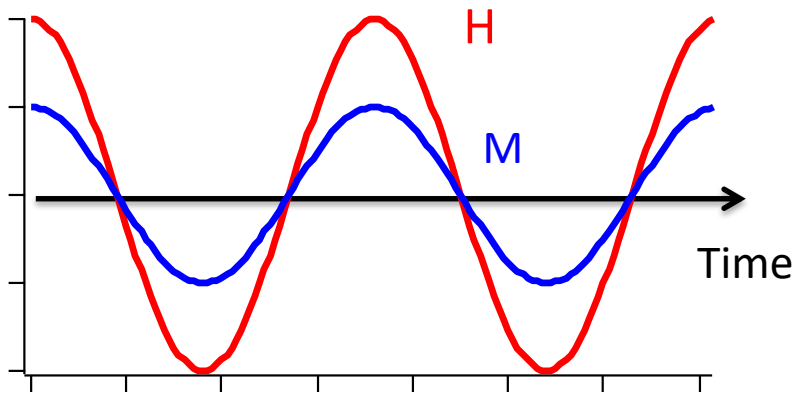
Power loss $\sim \Delta T$

FDA requirements: $H_{\max} f < 4.85 \times 10^8$ A-turns/m-s

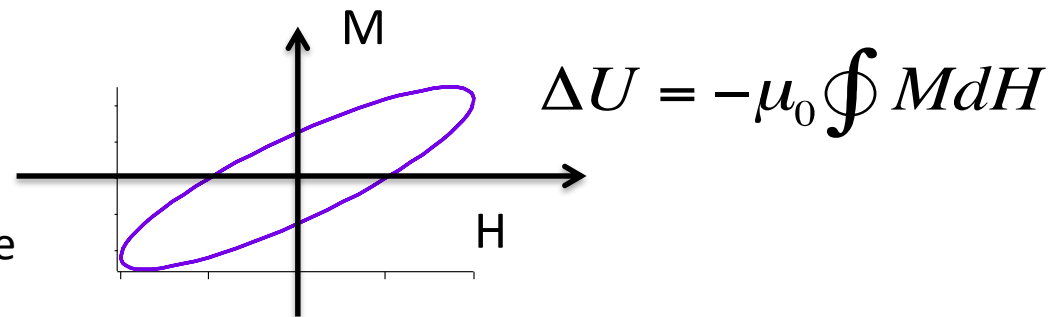
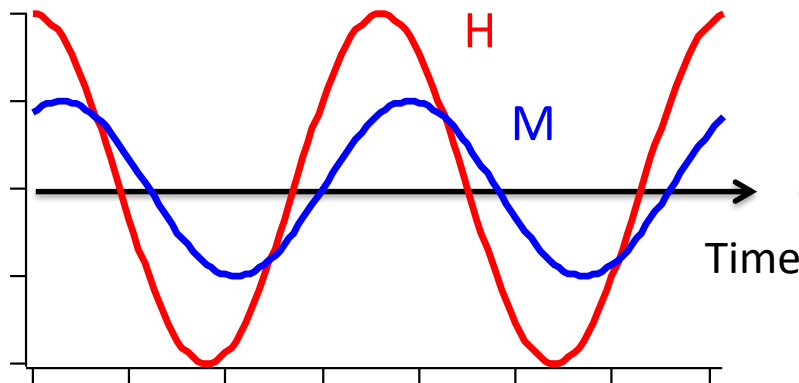
- with typical fields, $f_{\max} < 1.2$ MHz
- **Otherwise stimulate nerves and cause pain**

Magnetic Susceptibility $C = M/H$

Low amplitude AC field $H(t)$ drives magnetization $M(t)$

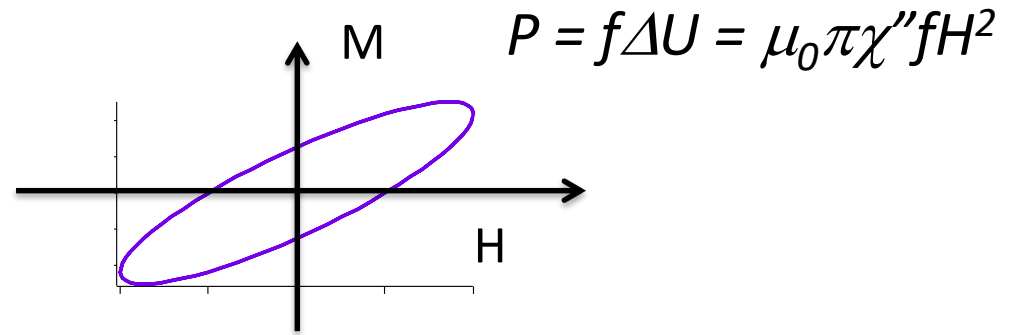
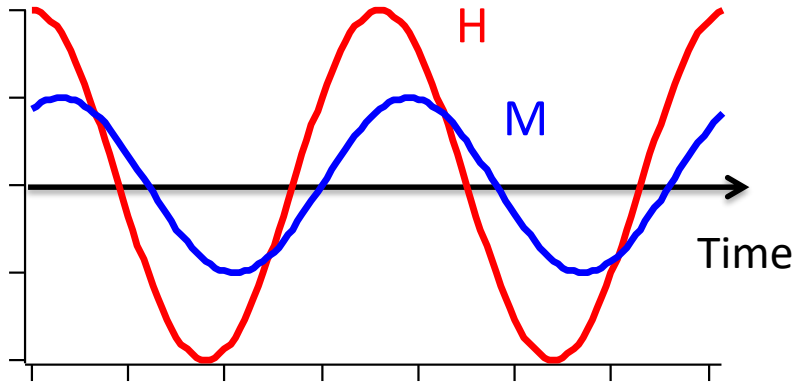


At higher frequency, $M(t)$ lags $H(t)$

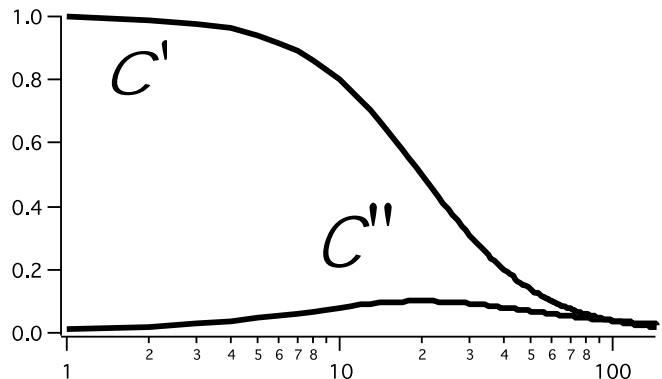


Hysteresis means energy dissipation

Imaginary Susceptibility



Can break $M(t)$ into components in-phase and 90° out-of-phase with $H(t)$, leading to the real and imaginary terms in the susceptibility, respectively



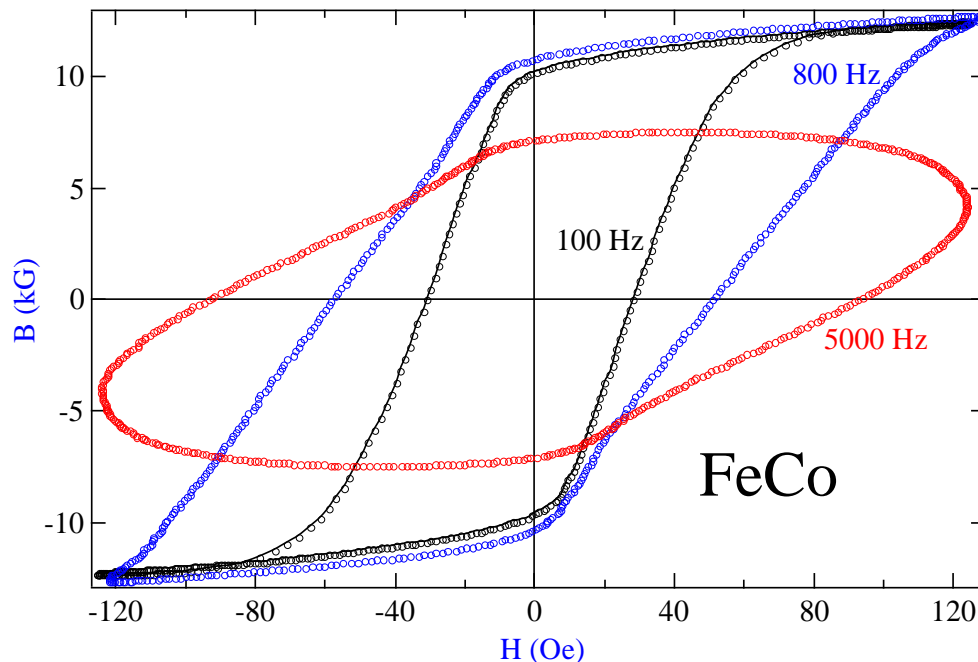
$$C = \frac{M}{H} = C' - iC''$$

Typical hyperthermia frequencies
100-500 kHz

Frequency

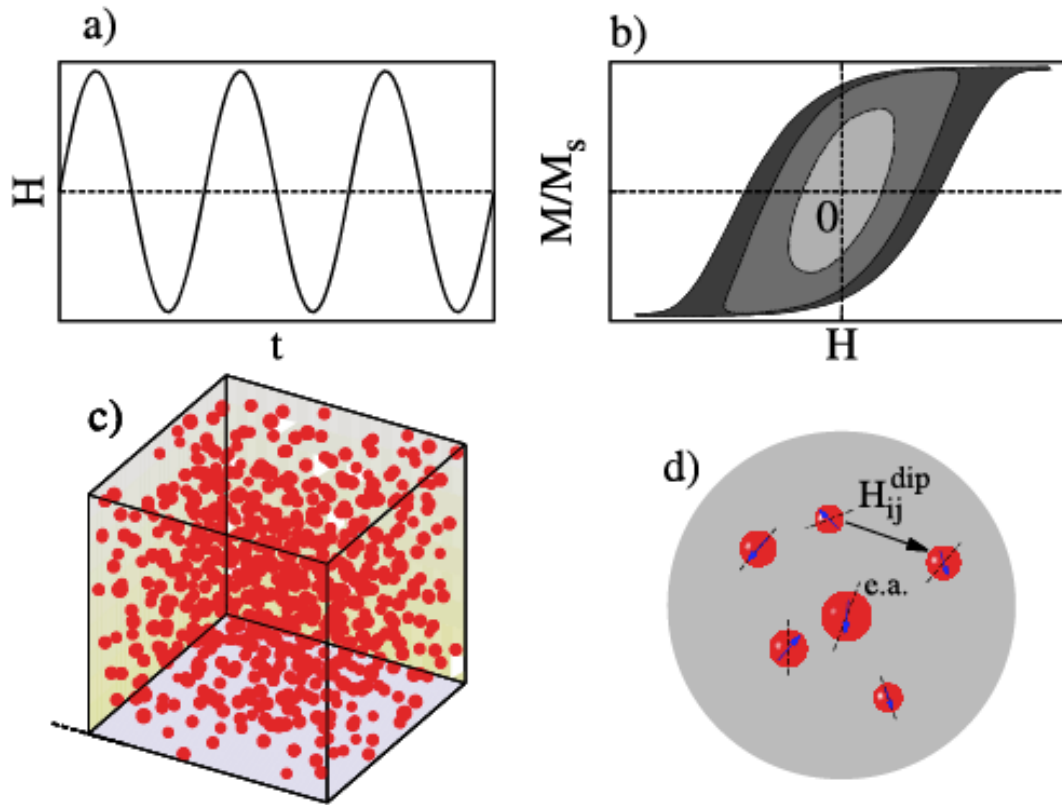
Are Superparamagnets Best?

- Many papers claim that SP particles are best for hyperthermia, but these particles have no magnetic energy losses
- Particles are SP in nearly DC measurements, but loops open up at AC frequencies used for hyperthermia

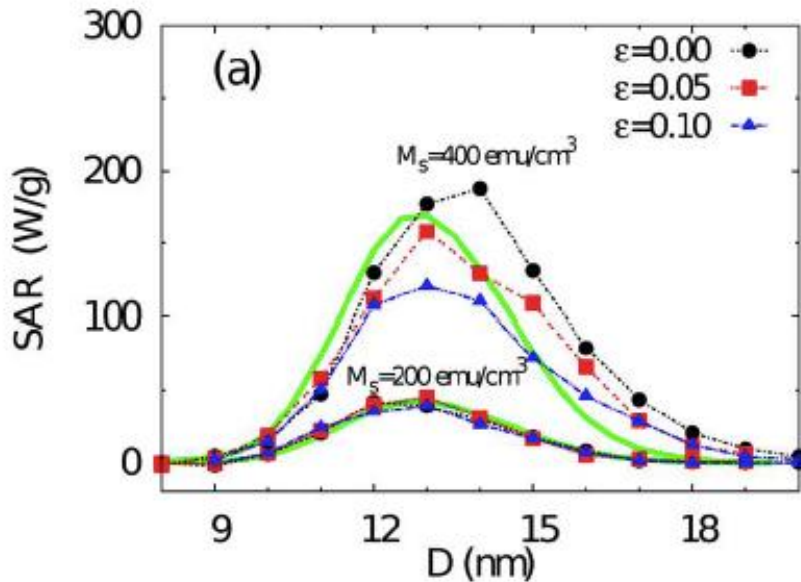


A. K. Giri, K. M. Chowdary, and S. A. Majetich, *Mater. Physics and Mechanics* **1**, 1-10 (2000).

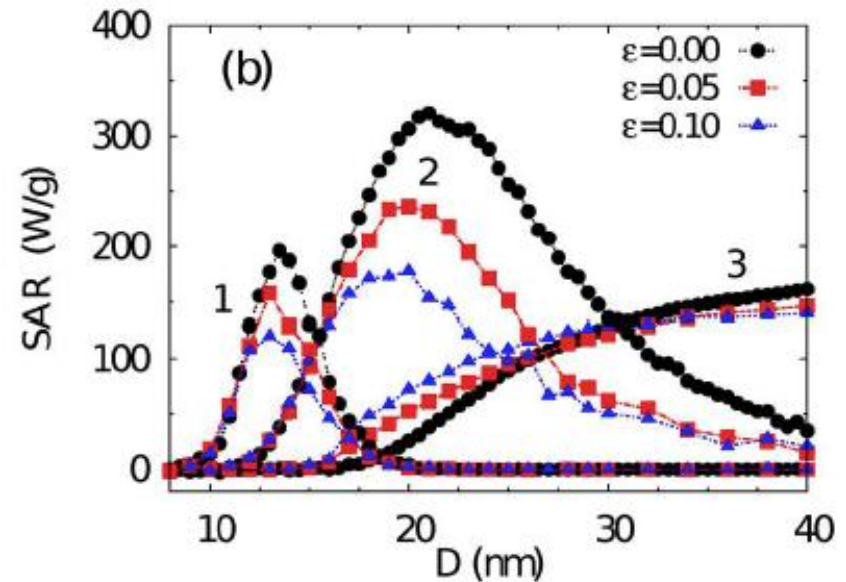
Hyperthermia Simulations



Hyperthermia Simulations: Magnetostatic Interactions and K



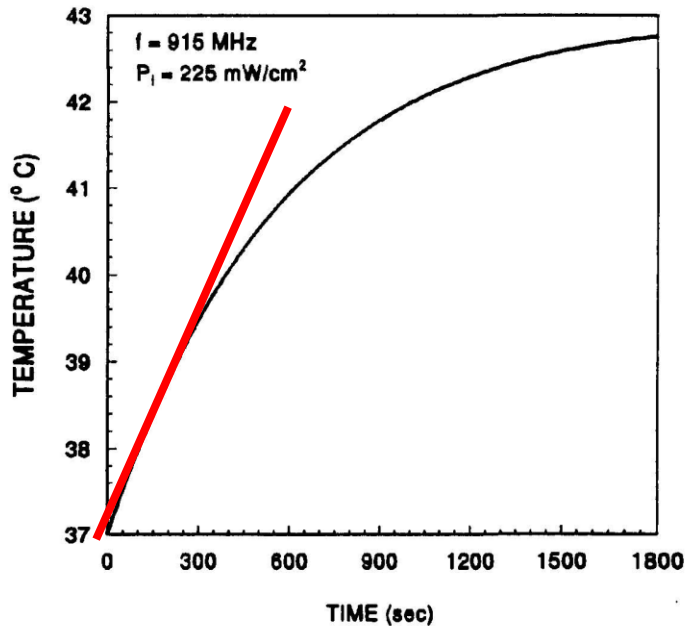
Vary packing fraction ε



Vary anisotropy

- 1: $3 \times 10^5 \text{ erg/cc}$
- 2: $1.5 \times 10^5 \text{ erg/cc}$
- 3: $0.5 \times 10^5 \text{ erg/cc}$

Quantifying Heating Power



Ex vivo measurement of heating rate

- measure initial slope of $T(t)$
- depends on particle concentration
- depends on whether particles immobilized (Brownian vs. Néel rotation)
- use IR T sensor, not thermocouple

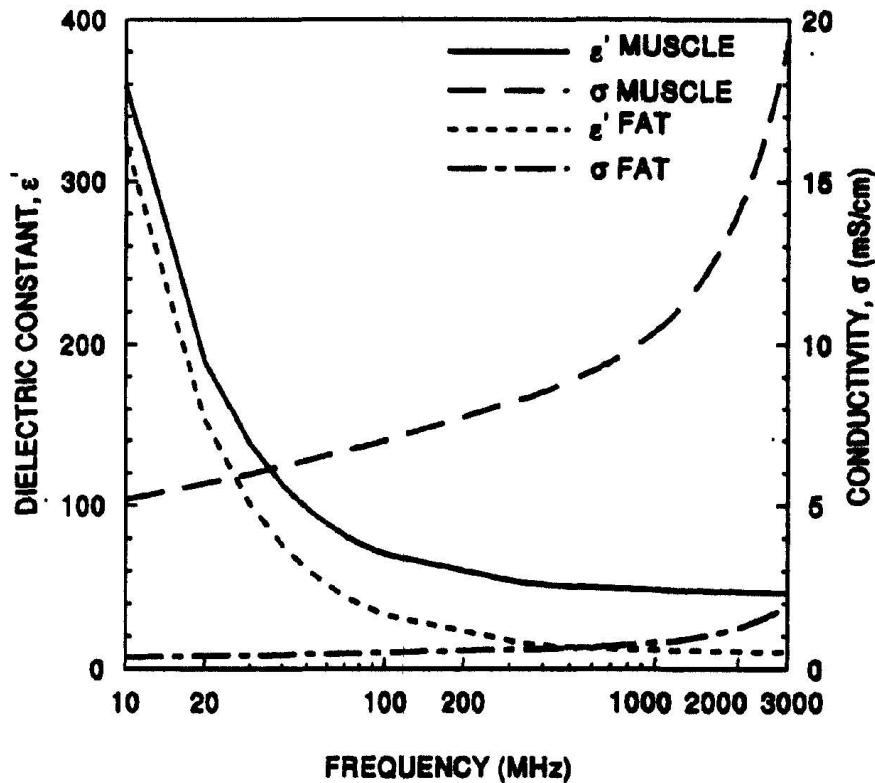
Figures of Merit

SAR – Specific absorption ratio – Power absorbed *per kg tissue*

SLP – Specific loss parameter – Power absorbed *per g Fe*

Thermal dose – *Should* be total power transferred, but not yet standardized

Electromagnetic Properties of Biological Tissues



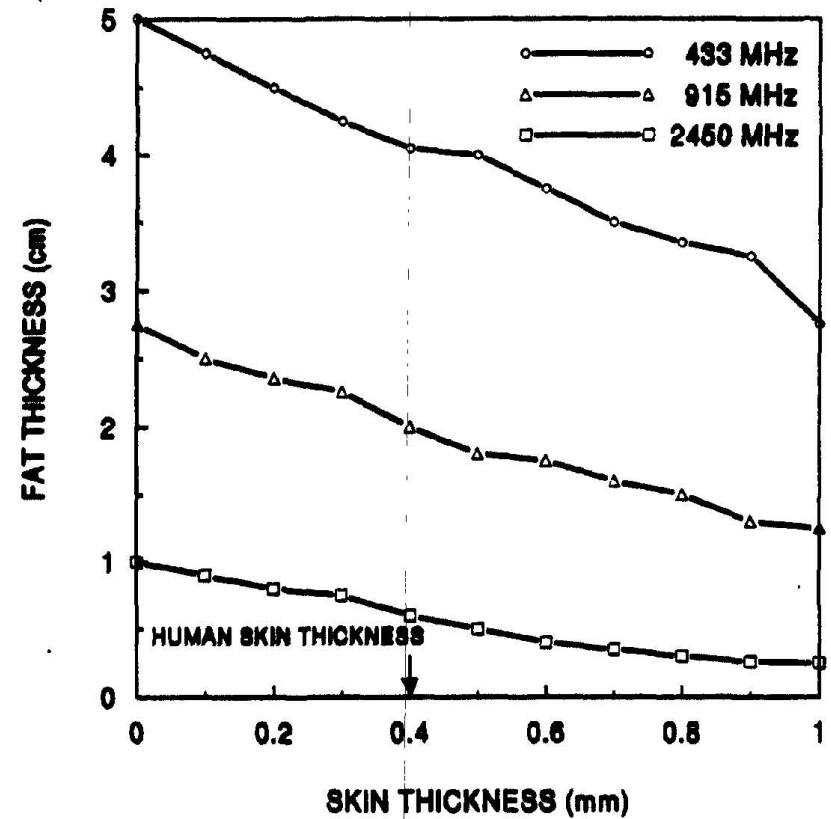
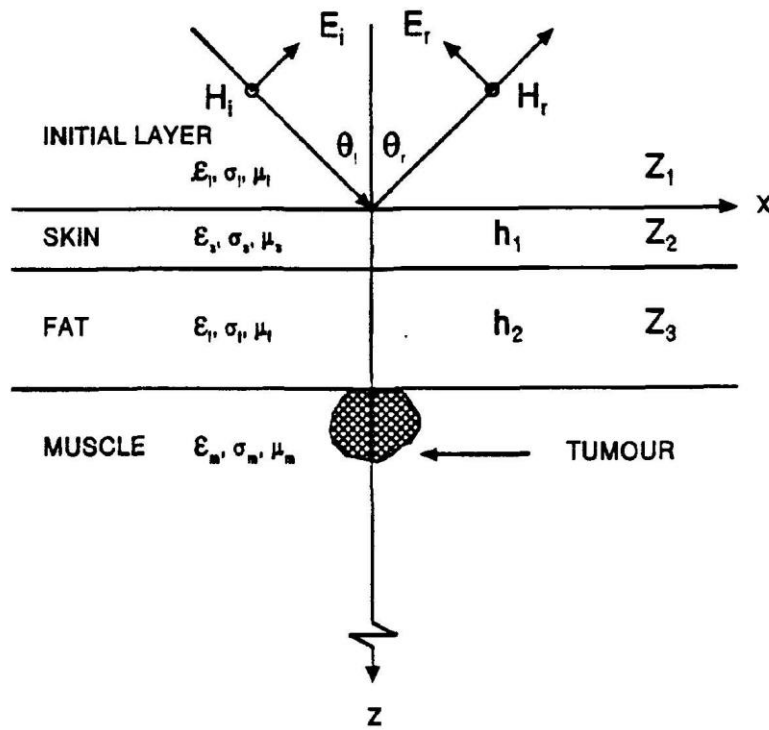
Low electrical conductivity at low frequencies, but not at high

- like a Capacitor

Muscle and fat have different electrical properties

- AC EM field hyperthermia can create hot spots in fat

Boundary Value Problems



EM field penetration depends on frequency, material, and thickness

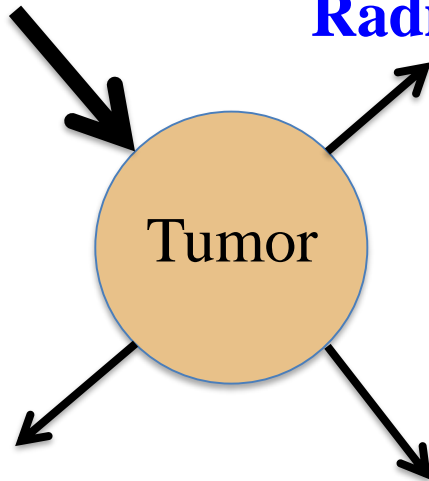
Must tailor hyperthermia treatment to individual

Bioheat Transfer Mechanisms

Power

Deposition

**Heat Convection,
Radiation**

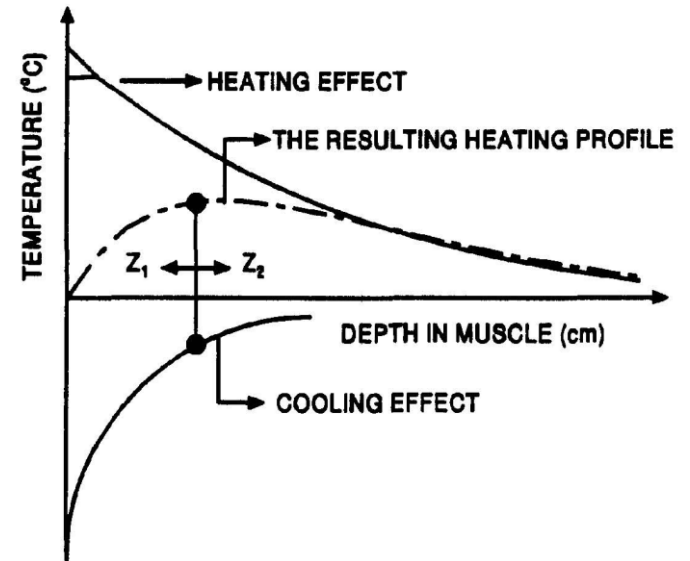


**Blood Perfusion,
Sweating**

Heat Conduction

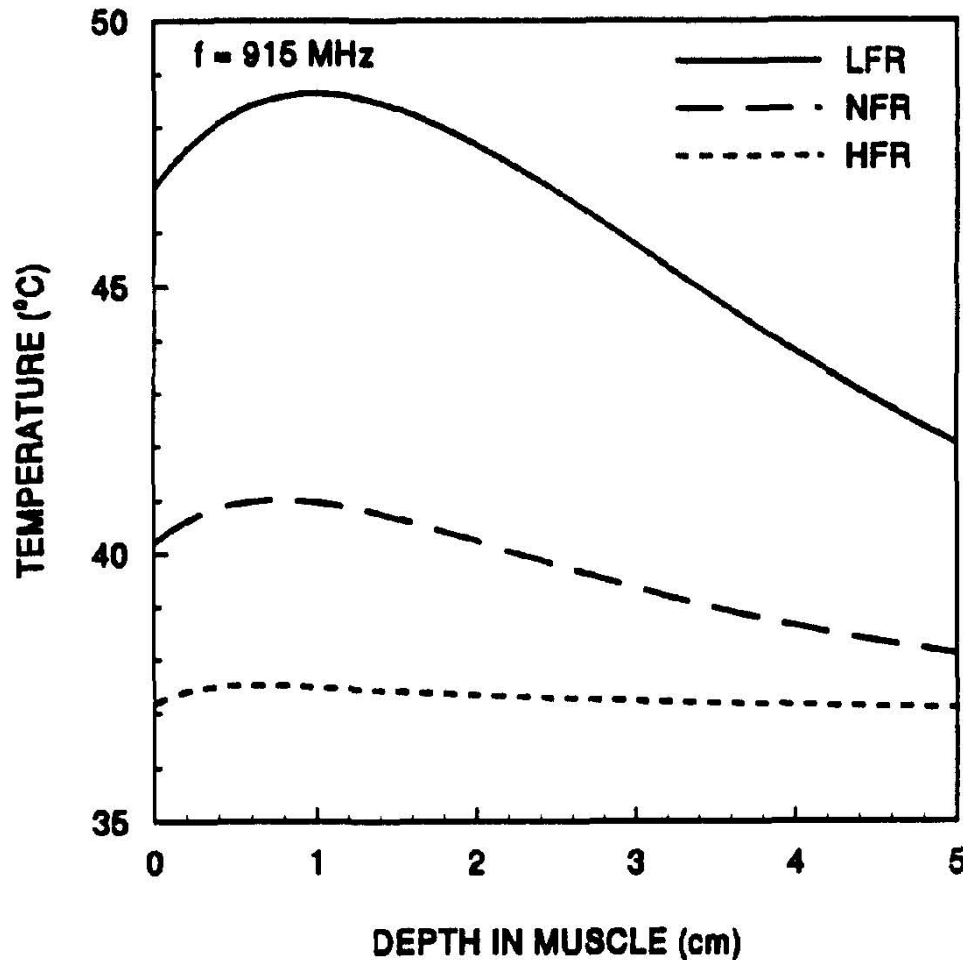
$$\rho c \frac{\partial T}{\partial t} = \nabla \cdot (k \nabla T) + P$$

Bioheat Equation



Don't want to damage
healthy tissue

Blood Perfusion



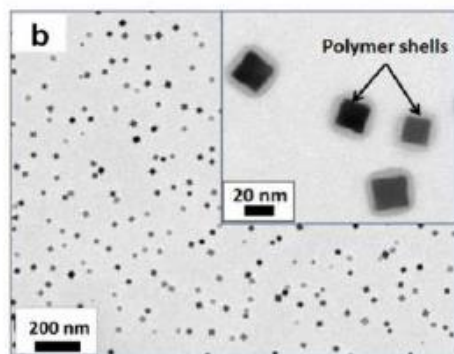
Low flow rate

Normal flow rate
(typical of muscle)

High flow rate (typical
of brain, kidneys)

Hyperthermia harder in
regions with high blood
flow rates

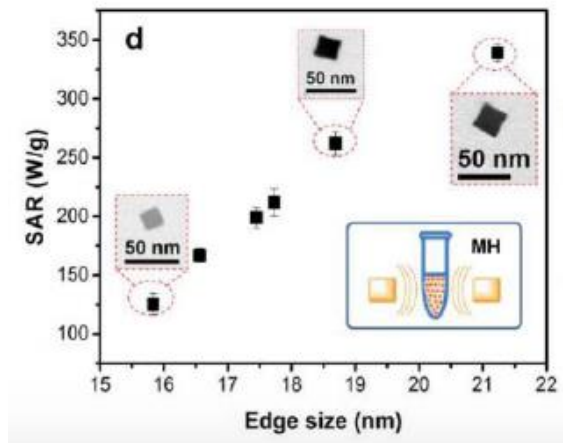
Nanocubes for Hyperthermia



17 – 21 nm Fe_3O_4 nanocubes currently have high SAR under conditions approved for in human application

Coatings to limit magnetostatic interactions are important

When particles taken up into endosomes, forming magnetosomes, they interact more strongly and the surface coating is often degraded. The stronger interactions reduce the SAR. This is also found with multicore NPs for hyperthermia.

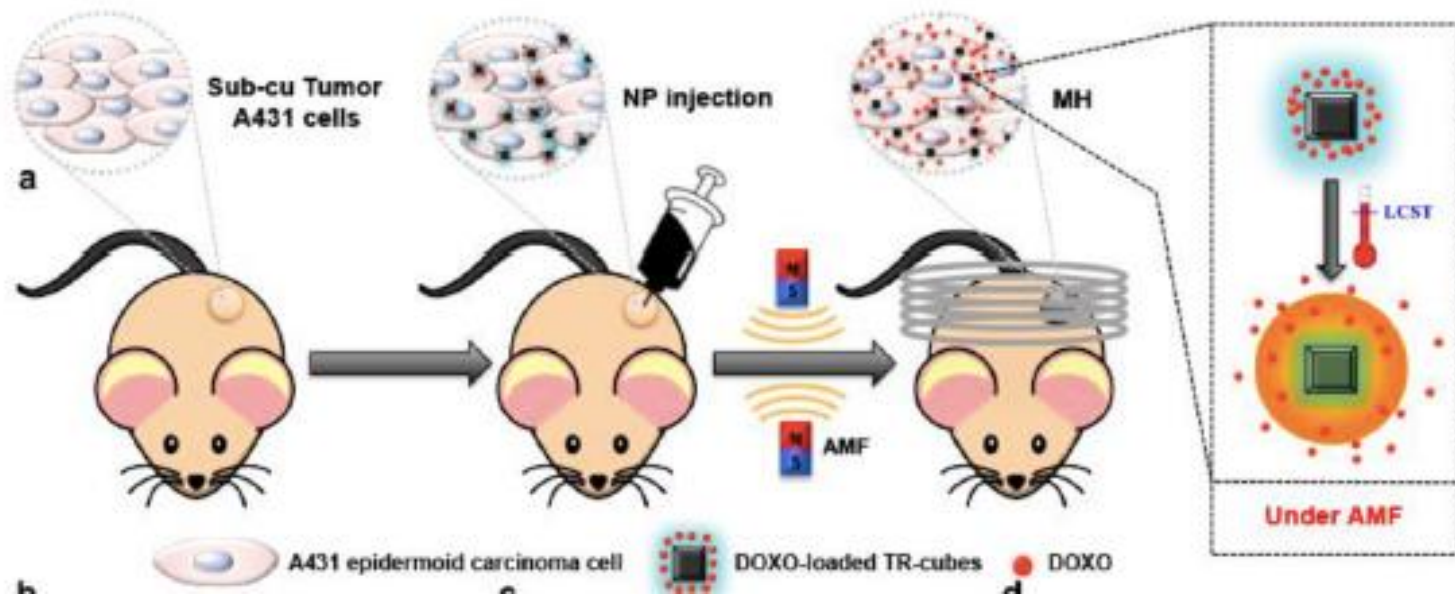


B. T. Mai, et al., ACS Appl. Mater. and Interf. **11**, 5727 (2019)

J. Kokosnjaj-Tabi, et al., ACS Nano **8**, 4268 (2014).

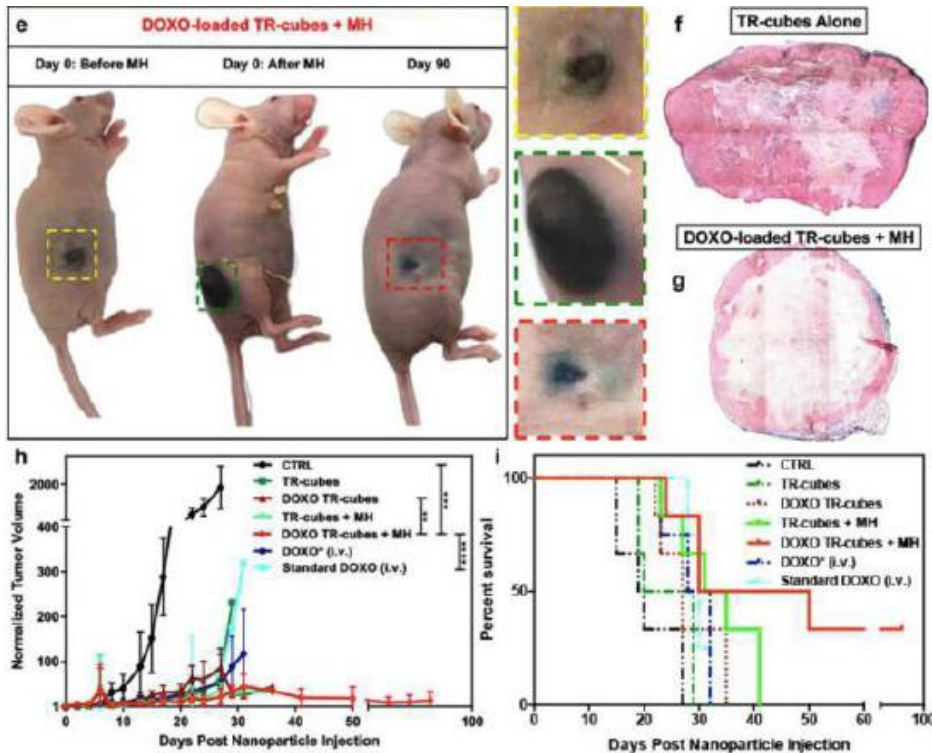
Nanocubes for Hyperthermia

AC magnetic field heats through the particles, and at the same time releases the chemo locally



Complete clearance of NPs after 5 months

Nanocubes for Hyperthermia



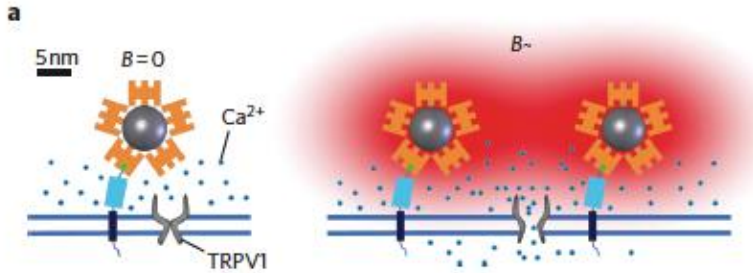
Hyperthermia plus localized drug release (doxorubicin) eliminates skin cancer tumors

DOXO has FDA approval for release through liposomes, and is used to treat breast, prostate, stomach, and colon

DOXO loaded in thermally responsive polymer coating the nanocubes

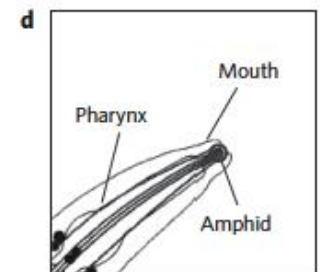
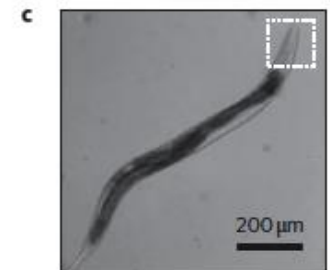
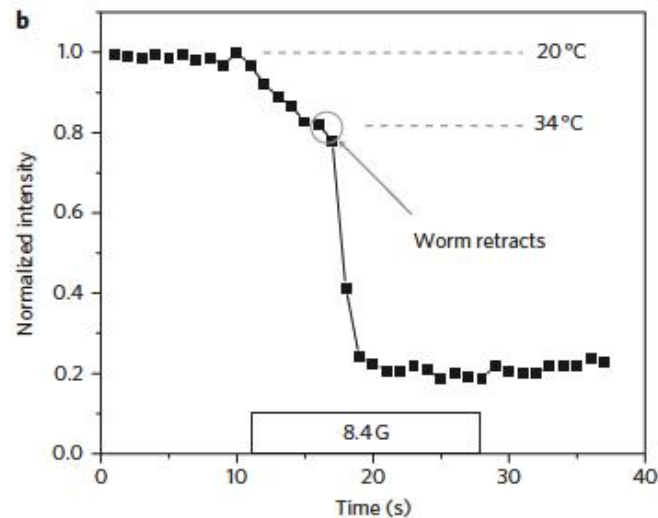
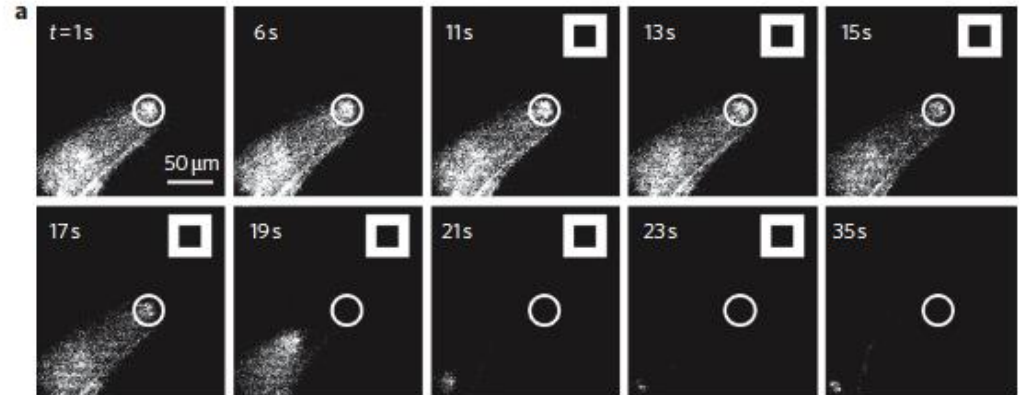
AC magnetic field heats through the particles, releases the chemo

Hyperthermia Control of Ion Channels



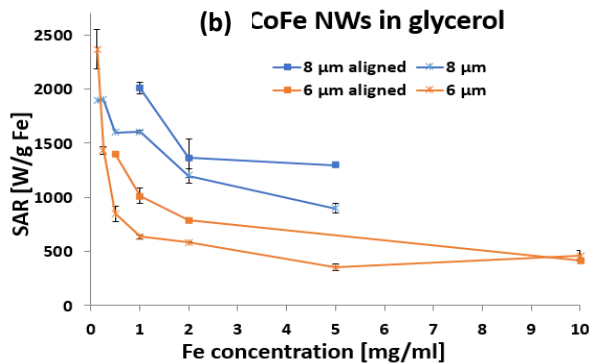
H. Huang, et al., Nat. Nanotech.
(2010), DOI:
10.1038/NNANO.2010.125

Use AC field to heat nanoparticles attached to sites near ion channels, see living *C. elegans* respond to AC magnetic field stimulus

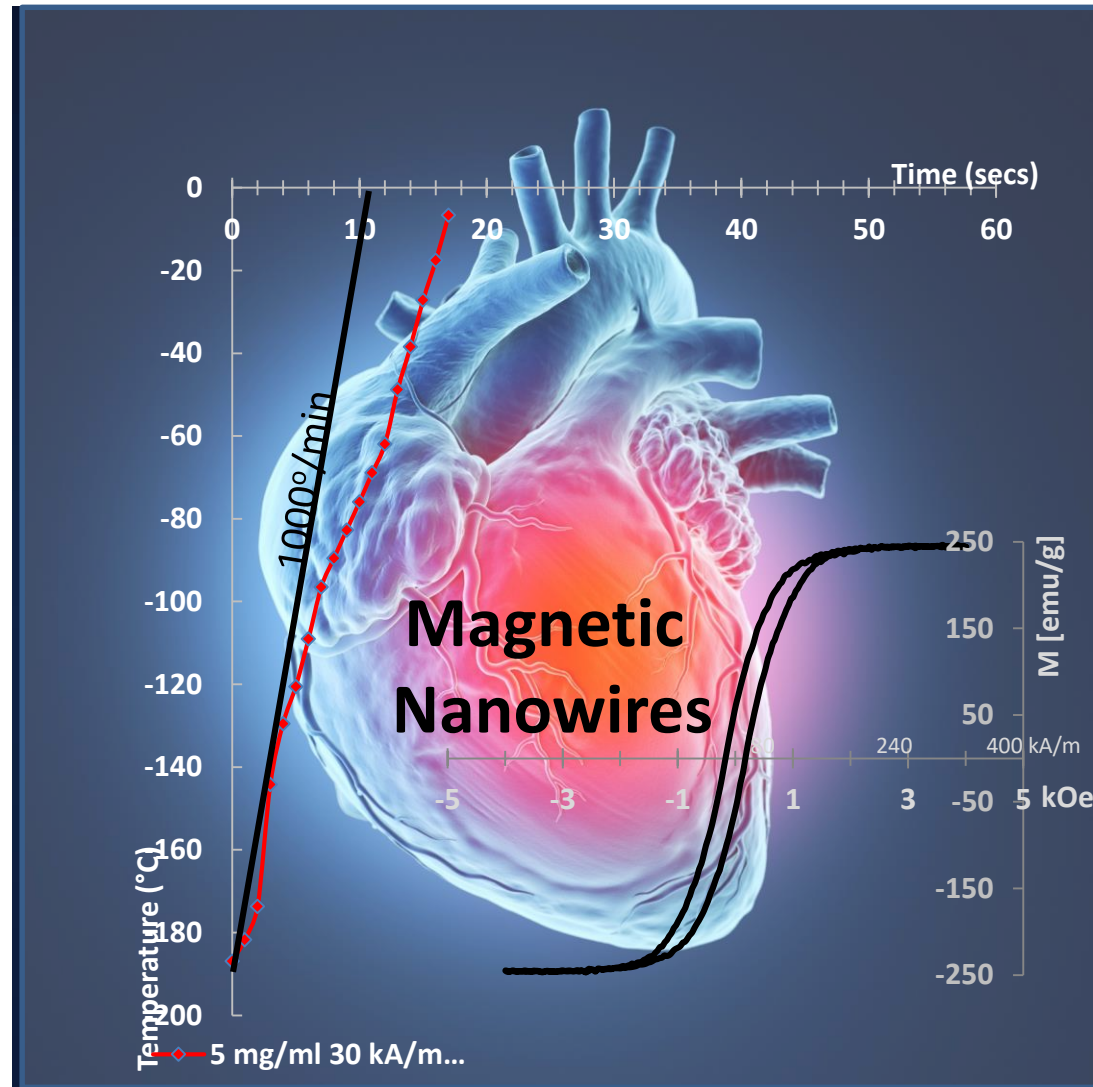


Hyperthermia for Organ Transplantation

The lifetime of harvested organs is extended by exchanging the blood with a liquid such as DMSO that doesn't crystallize when cooled. Rapid re-heating is needed prior to transplantation, or the glassy DMSO can recrystallize and damage the tissue.

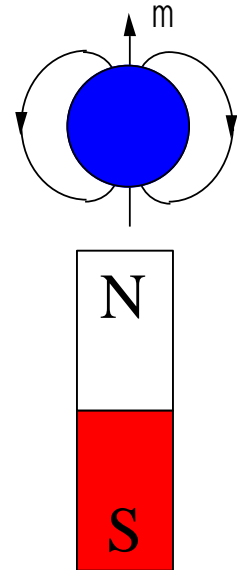


Beth Stadler and John Bischof, U. Minn.,
Oana Dragoa-Pinzaru, Iasi, Romania



Applications of Magnetic Nanoparticles

1. Intro and key nanoparticle properties
2. Magnetic manipulation
3. Magnetic sensing
4. Magnetic hyperthermia
5. Magnetic Particle Imaging (MPI)
6. Magnetic Resonance Imaging (MRI)



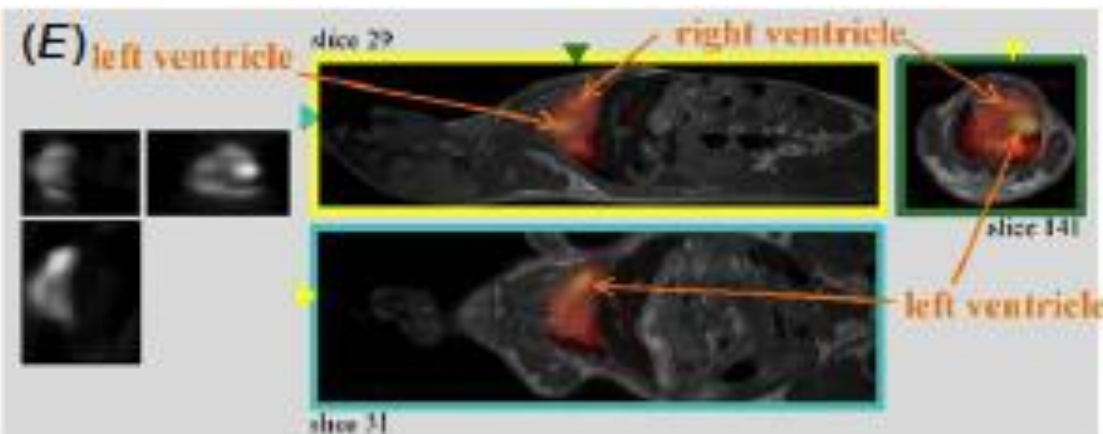
Magnetic Particle Imaging (MPI)

MRI – Detect protons (^1H) whose NMR frequency is perturbed by the field of a magnetic particle

MPI – Detect magnetic particles themselves

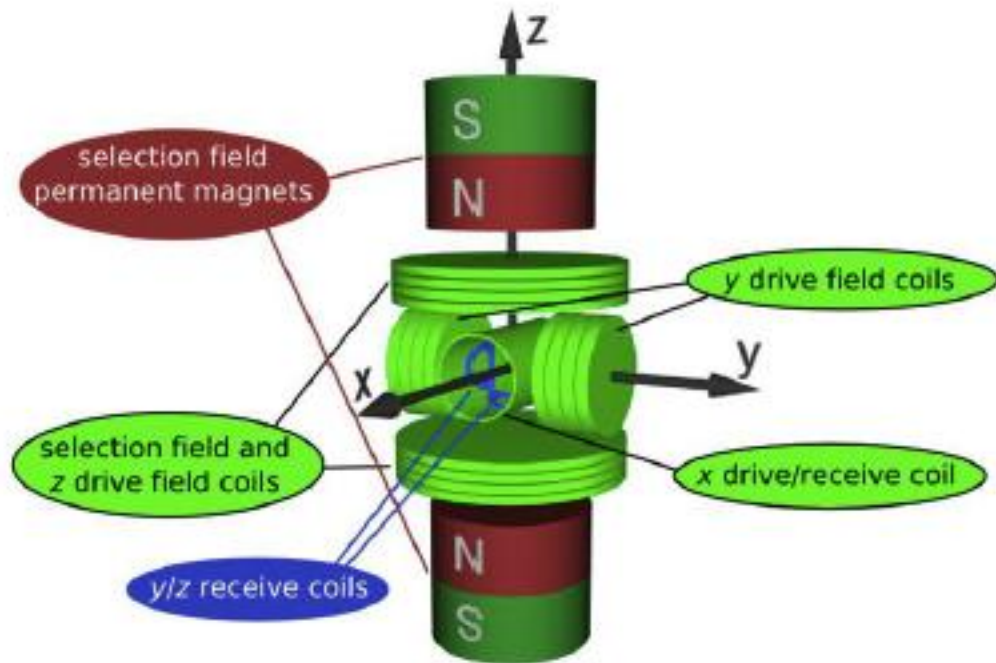
- 50 frames/s

- *see particles move* through mouse heart



B. Gleich, J. Weizenecker, Nature **435** 1214 (2005).

Magnetic Particle Imaging (MPI)



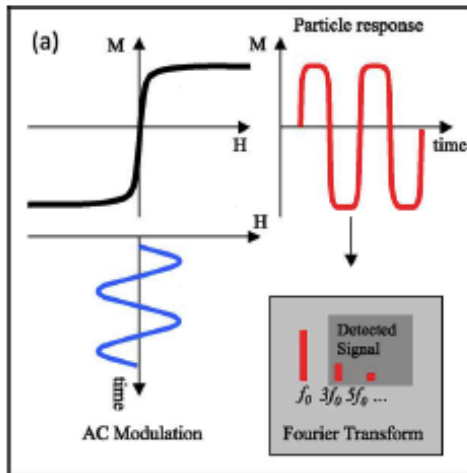
3 orthogonal DC fields determine Field Free Point (FFP)

Near $\mathbf{H} = \mathbf{0}$, NPs have a nonlinear response

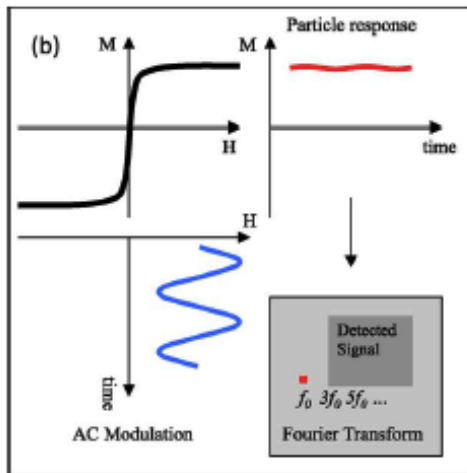
Apply high amplitude AC field, and response is dominated by NPs near the FFP

Scan FFP through sample and repeat to form 3D image

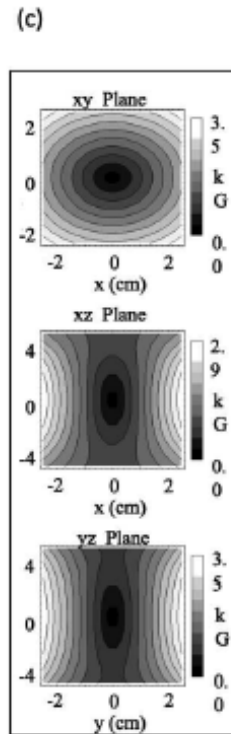
MPI – Nonlinear Response



a. Normal response



b. Saturated response




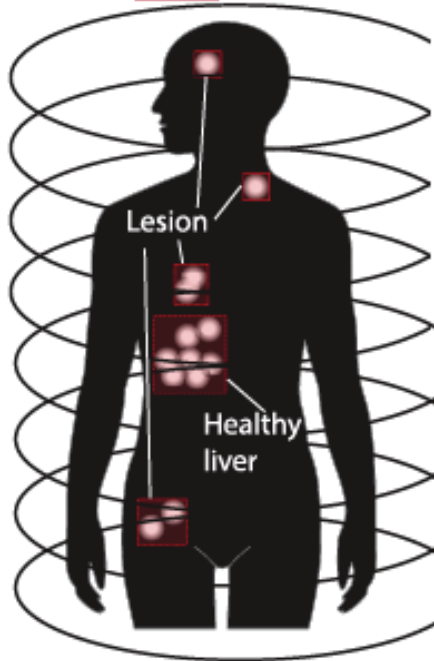
Excite with AC field about $H = 0$, FT has **high** amplitude in many harmonics

Excite about H near saturation, FT has **low** amplitude in many harmonics


Combining MPI and Hyperthermia for Deep Targeting

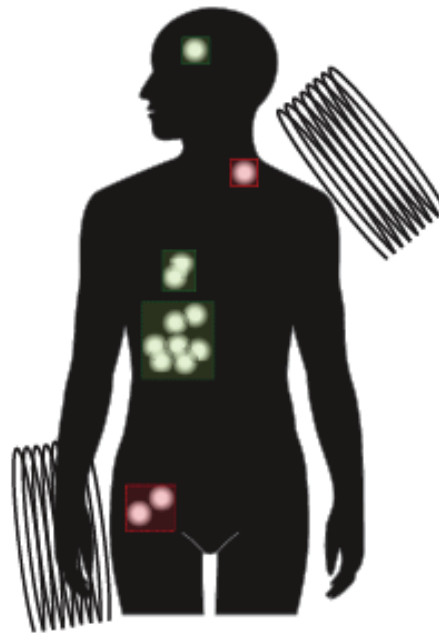
a Homogeneous AC coils target all tracer, including healthy sites of accumulation

 MFH heat deposition

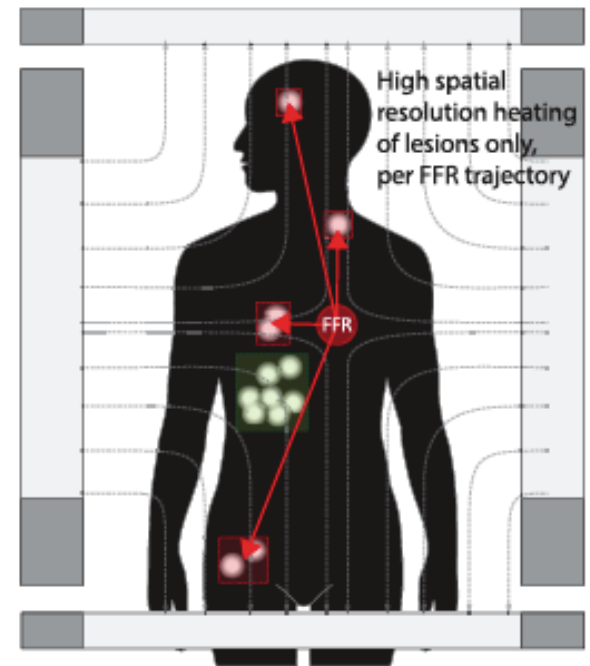


b Surface AC coils can target tracer near the surface only

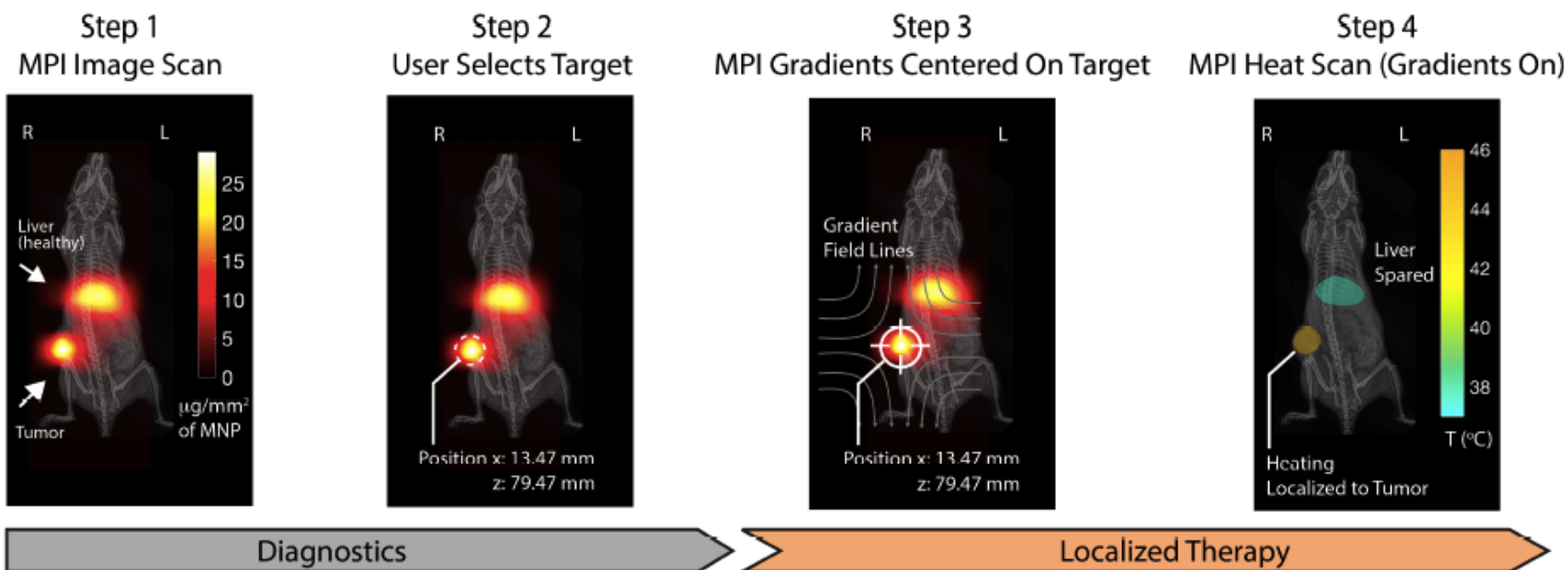
 No MFH heat deposition



c MPI-MFH can selectively target tracer anywhere, including deep in the body, while avoiding healthy sites of accumulation

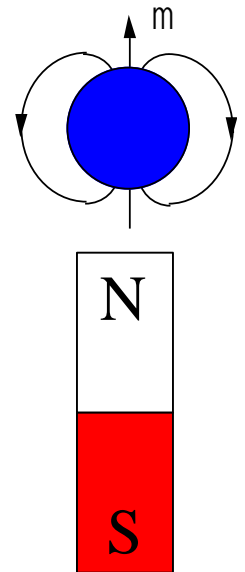


More MPI and Hyperthermia



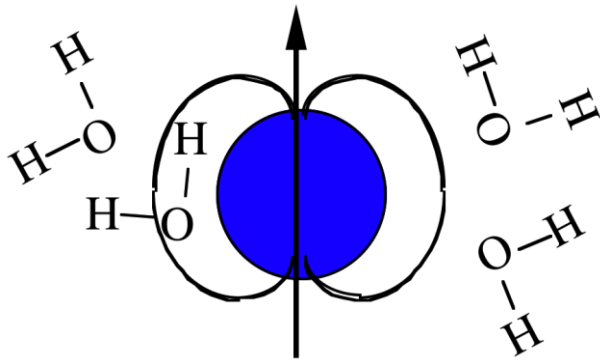
Applications of Magnetic Nanoparticles

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5. Magnetic Particle Imaging (MPI)
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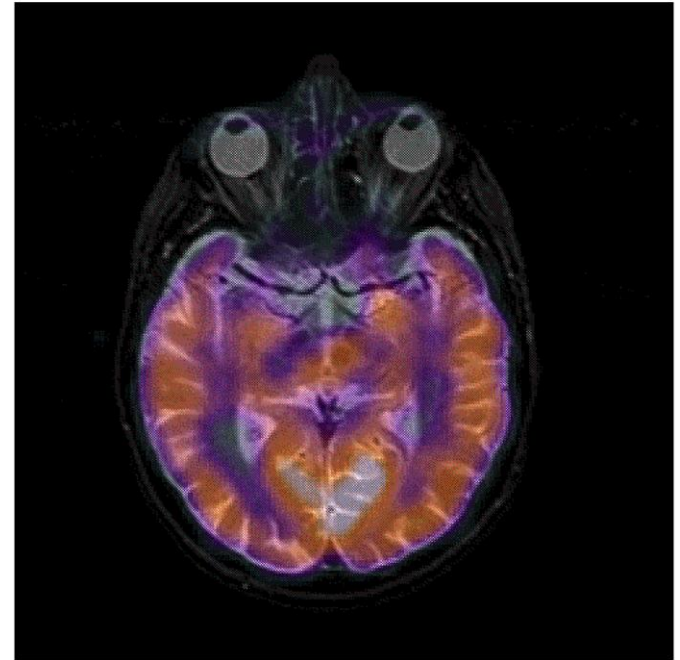
MRI CONTRAST AGENTS

**Particle moment generates a
Magnetic Field**



- Magnetic particle fringe field changes relaxation rate of nuclear moments of hydrogen atoms (¹H) in nearby water molecules

- Remove water background for higher contrast



Magnetic resonance imaging (MRI)

^1H Nuclear Magnetic Resonance (Proton NMR)

- Very sensitive method for detecting hydrogen nuclei in different chemical (and magnetic) environments
- **Nuclear spins** act like paramagnets with high saturation field
 - At DC field of 1.5 T, only *slight* imbalance
 - ^1H magnetic field is very weak, so **detect by dynamic measurements**
 - FMR typically in GHz range, ^1H NMR **typically in MHz range**
- Magnetic resonance imaging (MRI) = spatial mapping of NMR signal

Longitudinal and Transverse Relaxation

m precesses about a static field B_0

Apply rf field pulse to rotate by $\pi/2$

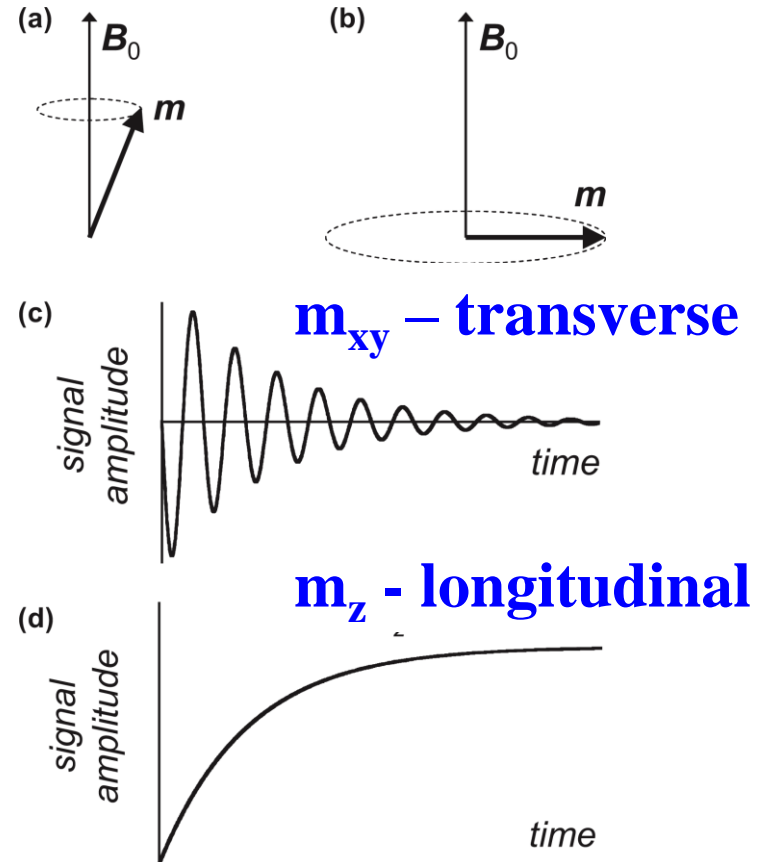
m precesses back to lie parallel to B_0

Bloch-Bloembergen equations

$$\frac{\partial m_z}{\partial t} = -g(\vec{m} \times \vec{B})_z = \frac{m_s - m_z}{T_1}$$

$$\frac{\partial m_{x,y}}{\partial t} = -g(\vec{m} \times \vec{B})_{x,y} = \frac{m_s - m_{x,y}}{T_2}$$

2 characteristic times, T_1 and T_2



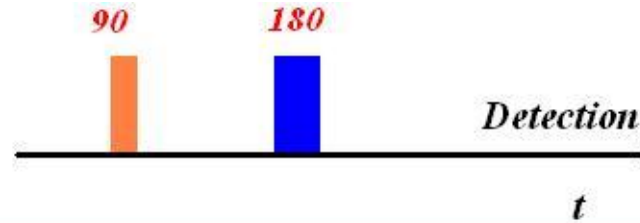
Q. A. Pankhurst, et al., J. Phys. D **36**, R167 (2003)

Dephasing and T_2

Spin Echo rf pulse sequence

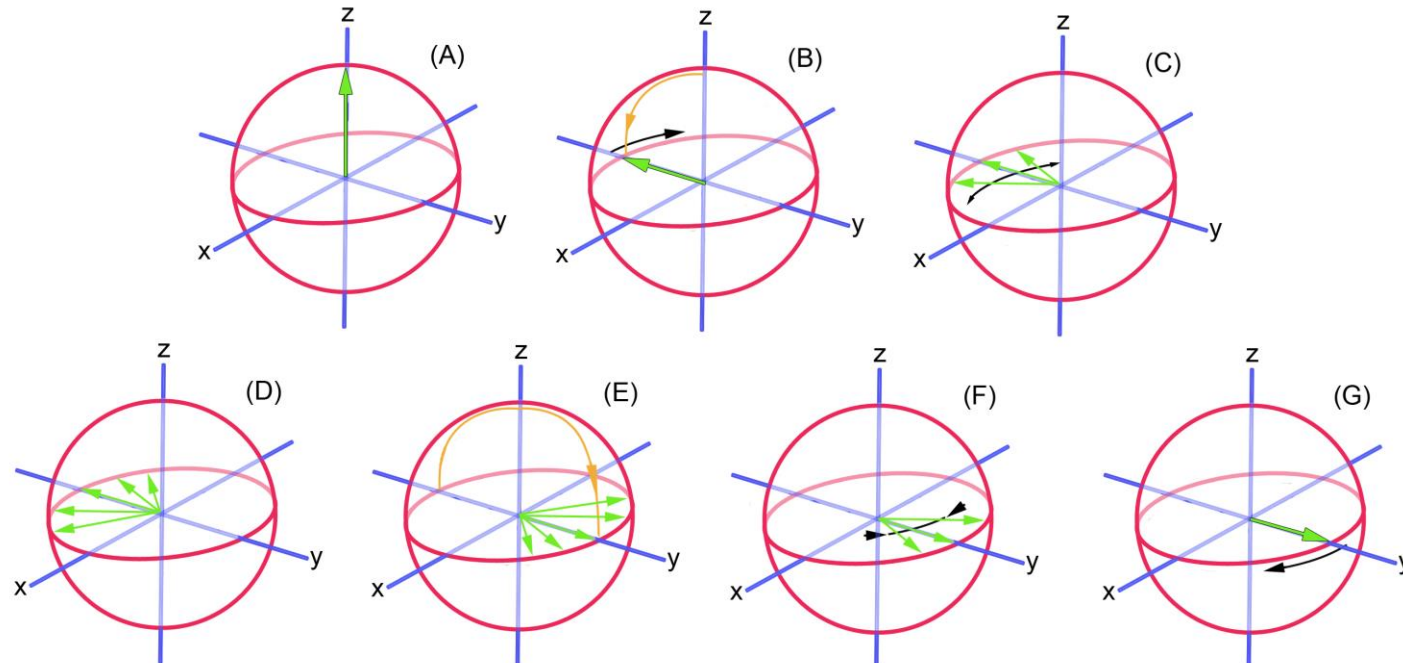
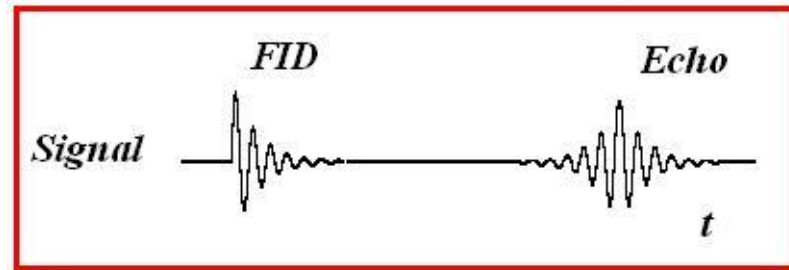
$\pi/2$ rotation

π rotation



$$\vec{B}_{tot} = \vec{B}_0 + \vec{B}_{rf} + \vec{B}_{local}$$

NPs change B_{local} , affect dephasing rate



$$W = gB_{eff}$$

T₁ and T₂ Relaxation

$$m_{x,y} \propto \exp(-t/T_2)$$

Decrease T₂ of water protons and signal decreases – **image gets darker**

- **Use iron oxide nanoparticles**

$$m_z \propto [1 - \exp(-t/T_1)]$$

Decrease T₁ of water protons and signal increases – **image gets brighter**

- **Use Gd³⁺DTPA**

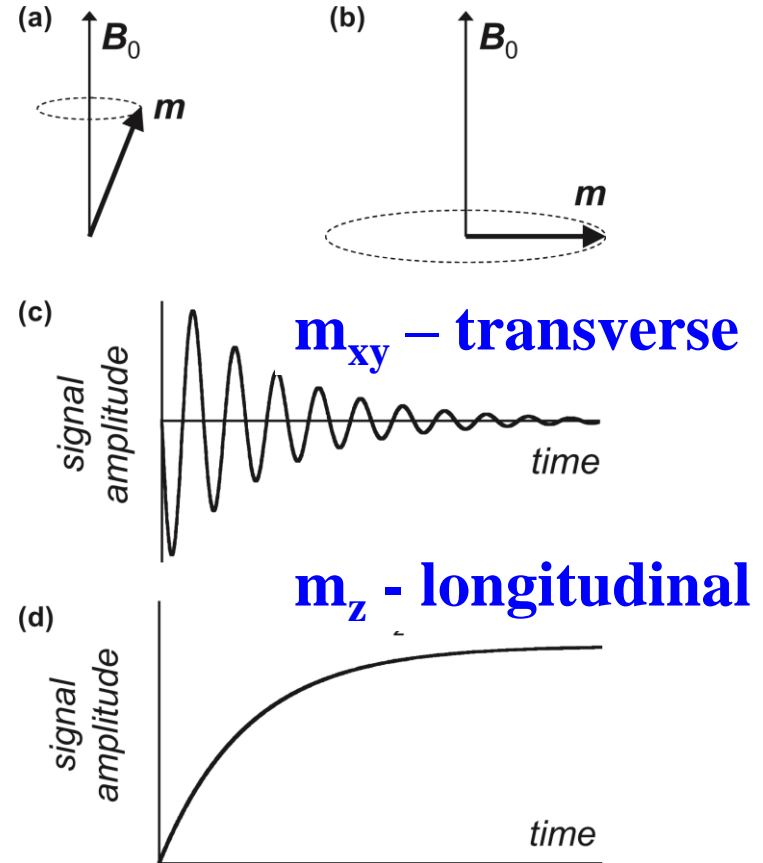


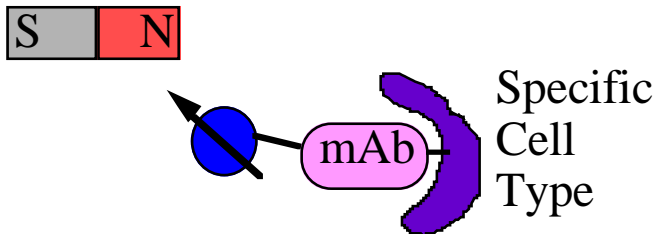
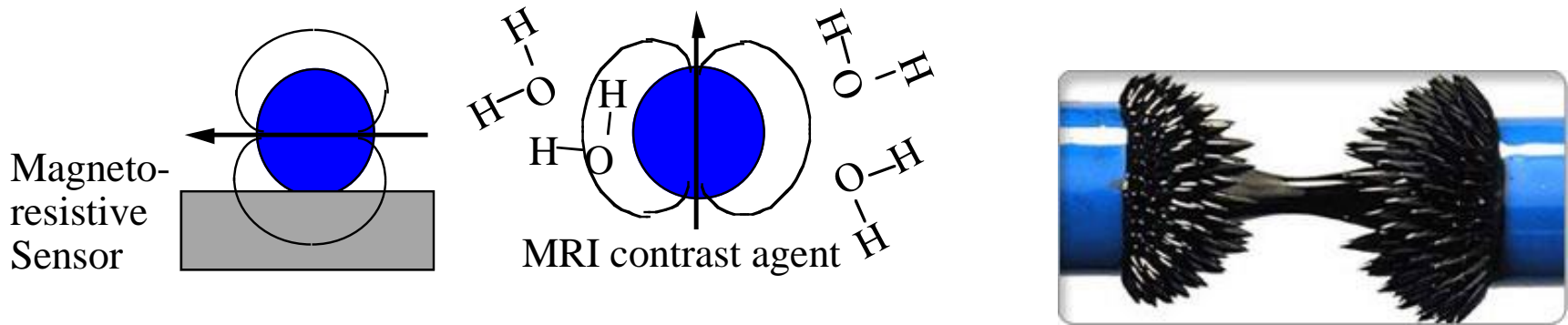
Figure of Merit:

$$W = gB_{eff}$$

Relaxivity R₁ (or R₂) = change in T₁ (or T₂) per concentration of contrast agent

Summary

Most magnetic particle applications arise because they can generate magnetic fields, can move or change properties in response to a magnetic field, and can easily be bound to other things



Acknowledgments



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Washington

Teresa Pellegrino – U. Genoa

JitKang Lim – U. Sains Malaysia

Beth Stadler – U. Minnesota